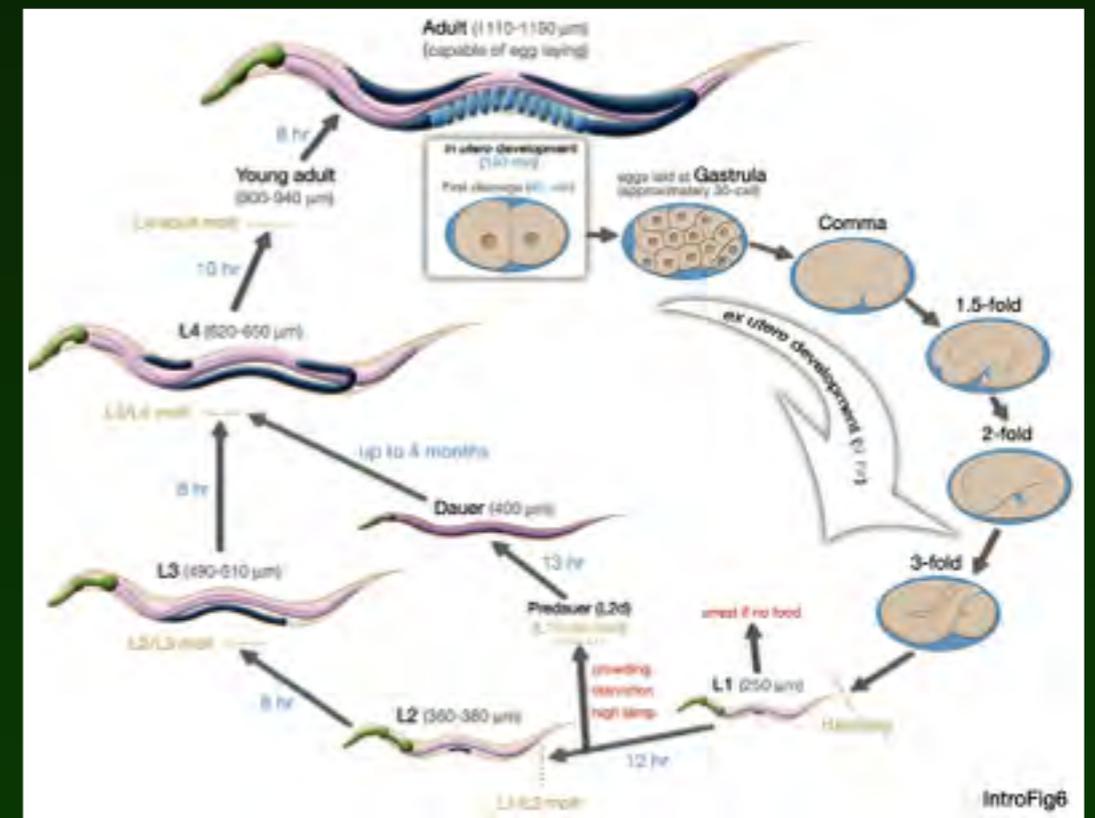


Noises and dynamics in cells: Mathematical modeling in systems biology

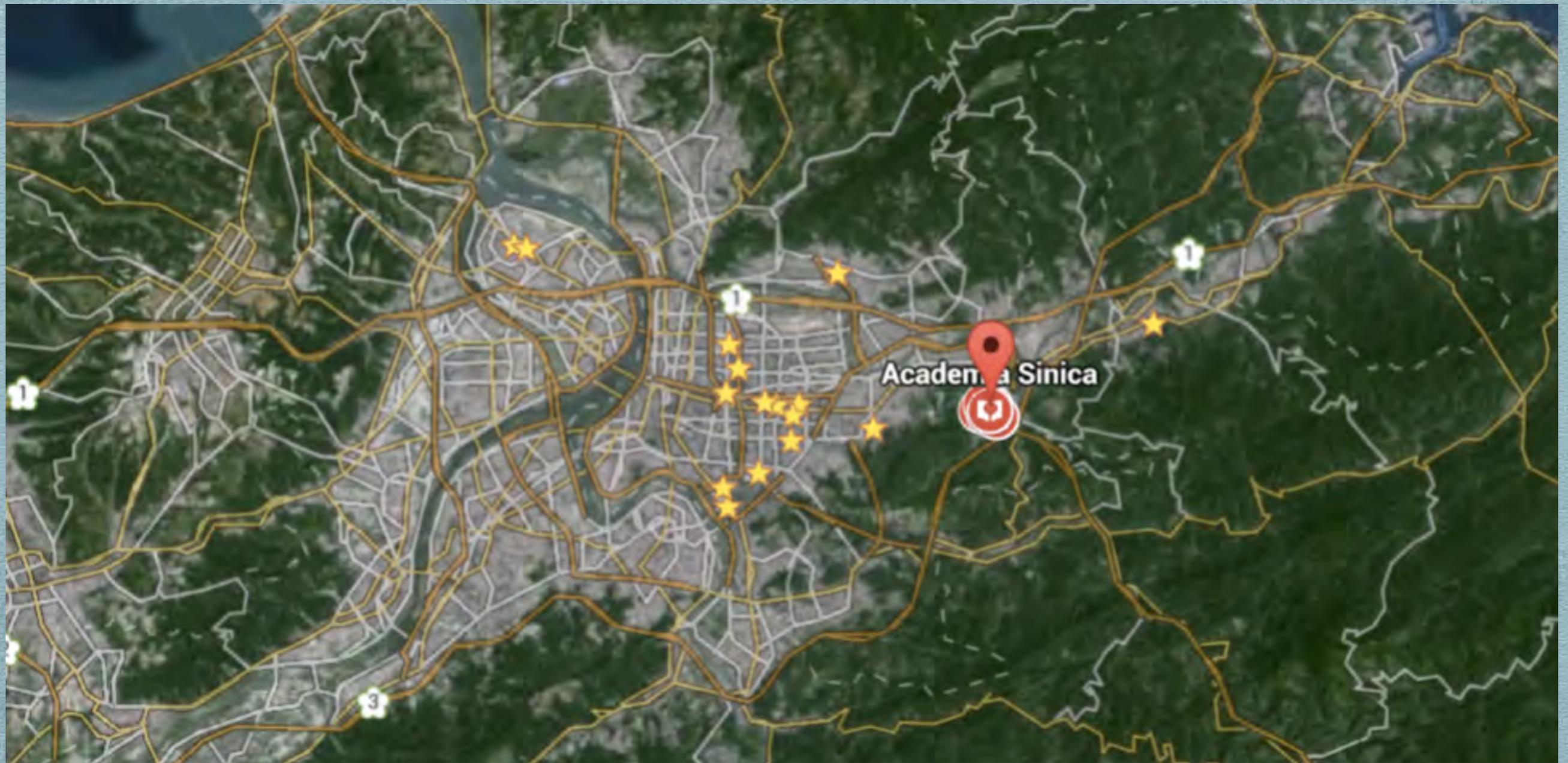
Chao-Ping Hsu
Inst. of Chem., Academia Sinica
中央研究院 化學所 許昭萍

Webinar
Philippine Genome Center
May 23, 2016





Where am I?



Academia Sinica is located to
the East of Taipei City



Institute of Chemistry
Academia Sinica

My research interests

- Electron and energy transfer problems.
 - In quantum chemistry.
 - With an application to **photosynthesis**.
 - most of our application are in Materials.
- Dynamical modeling in Biology.

Hierarchy in Biology

A multi-cellular organism

Organs

Tissues

Cells

Organelles

Molecules

Building units

On the other hand, "systems" level of understanding based on molecular studies have started to emerge.

A lot of work in Chemistry and Biology aim to understand the structure and function at molecular level.

Proteins, DNAs,
Lipids, Glycoses

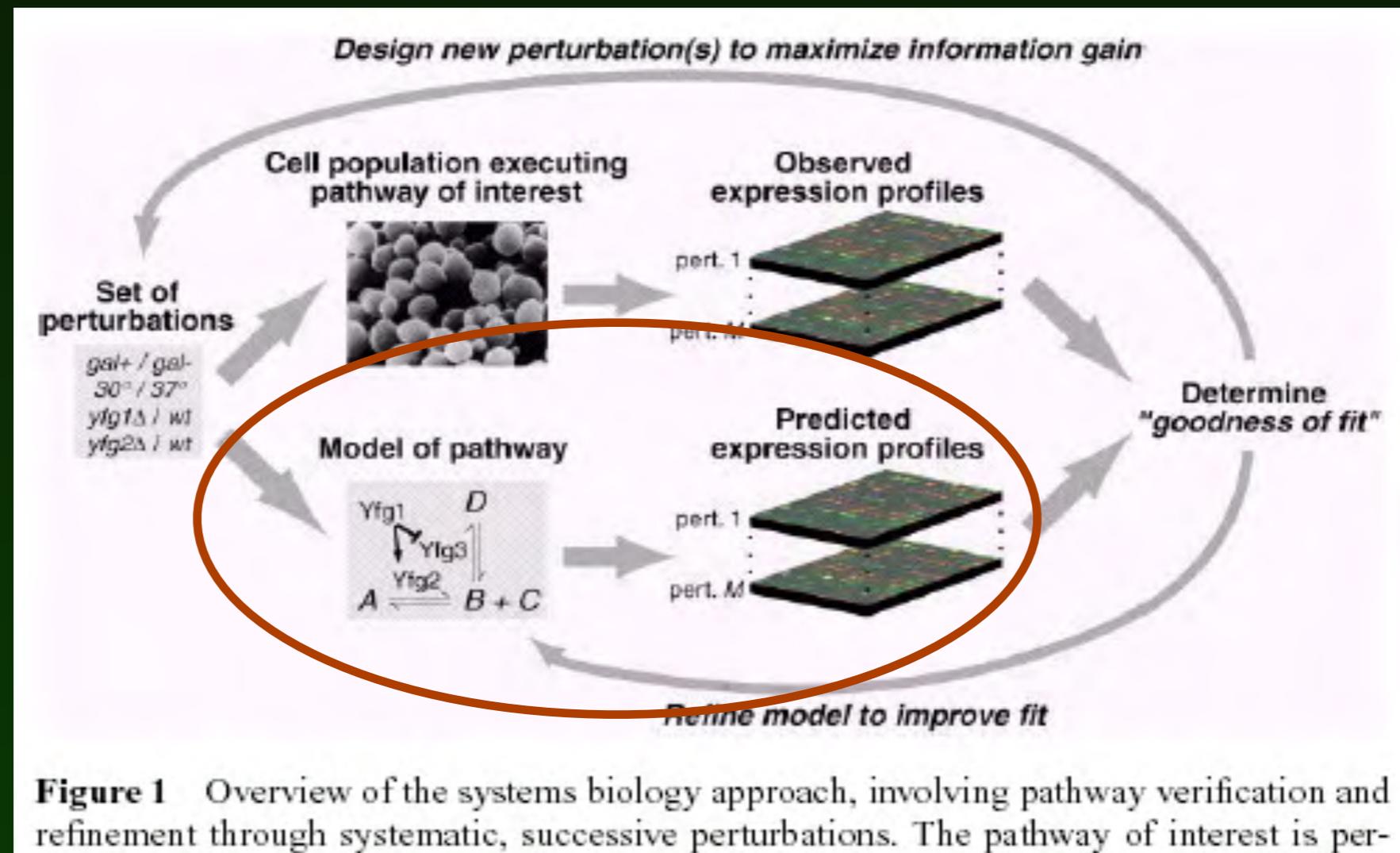
Amino Acids
Nitrogenous Bases

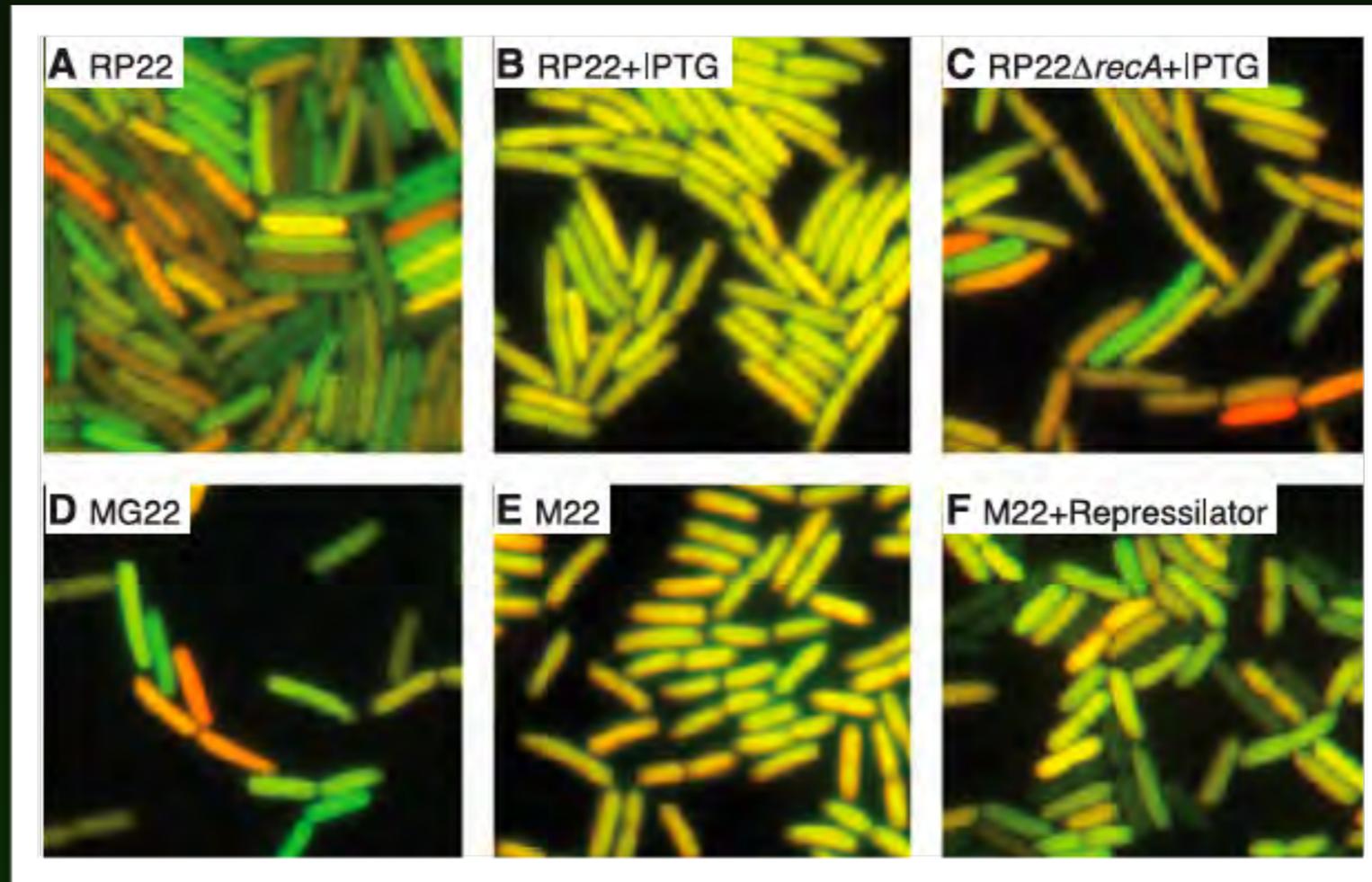


We are interested in

- Dynamic Description of Biological systems.
- What is really going on in a living system?
 - How the desired outputs are generated.
 - What may be the factors that contribute to the special properties of such dynamics.
 - ...

In the framework of biological observations



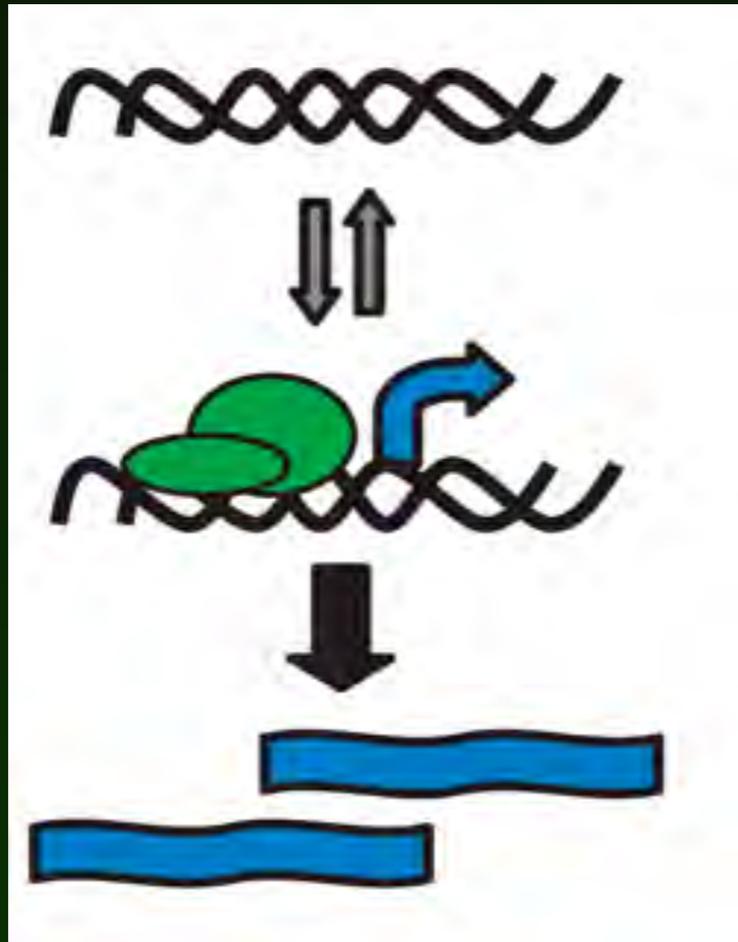


Elowitz, 2002

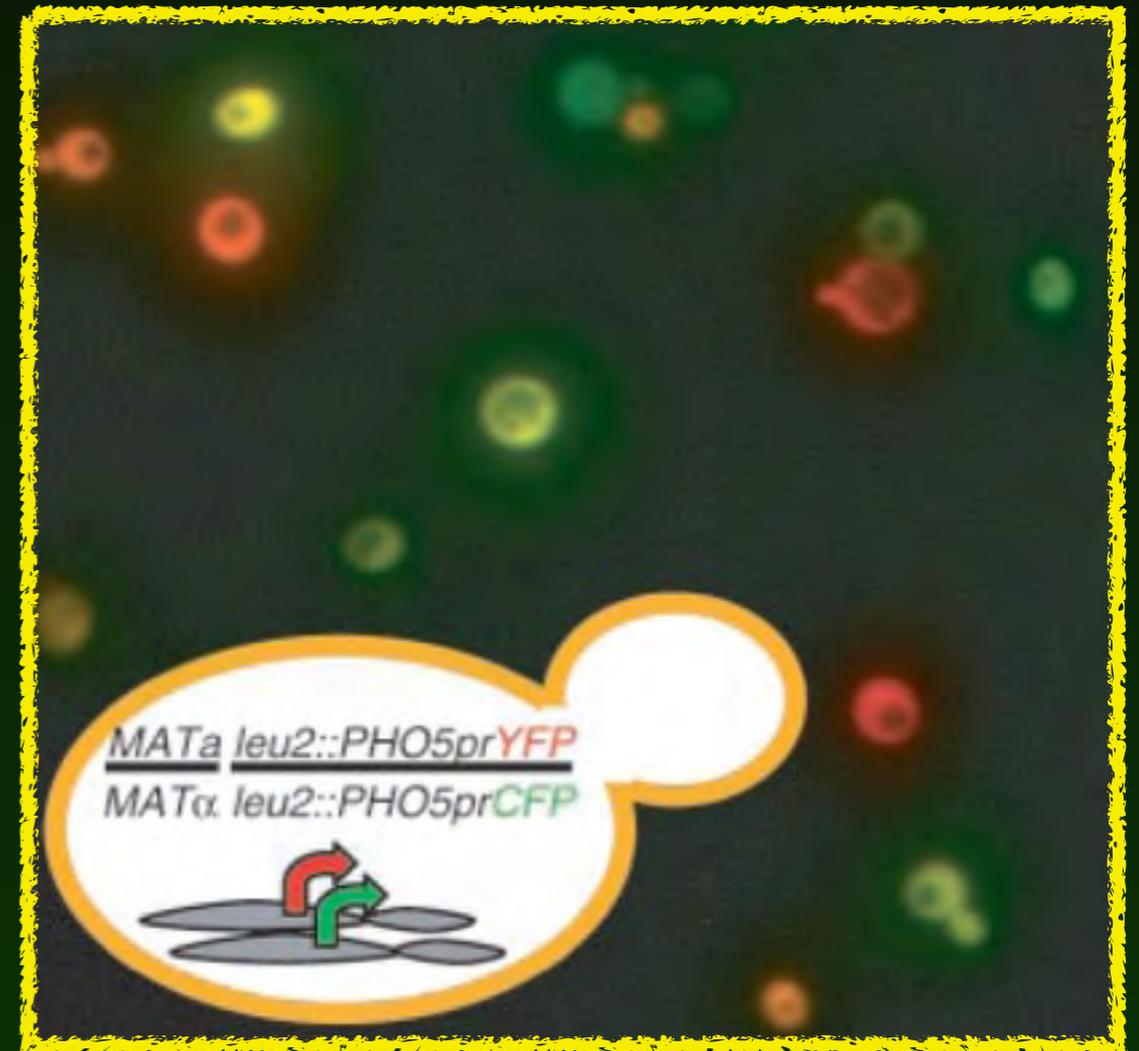
Dynamics in stochastic biological processes — “noisy cells”

Gene expression is "noisy"

inactive



active



Gene transitions randomly between active and inactive states

Cell to cell variability in a population

Science 309, 2010-2013 (2009)

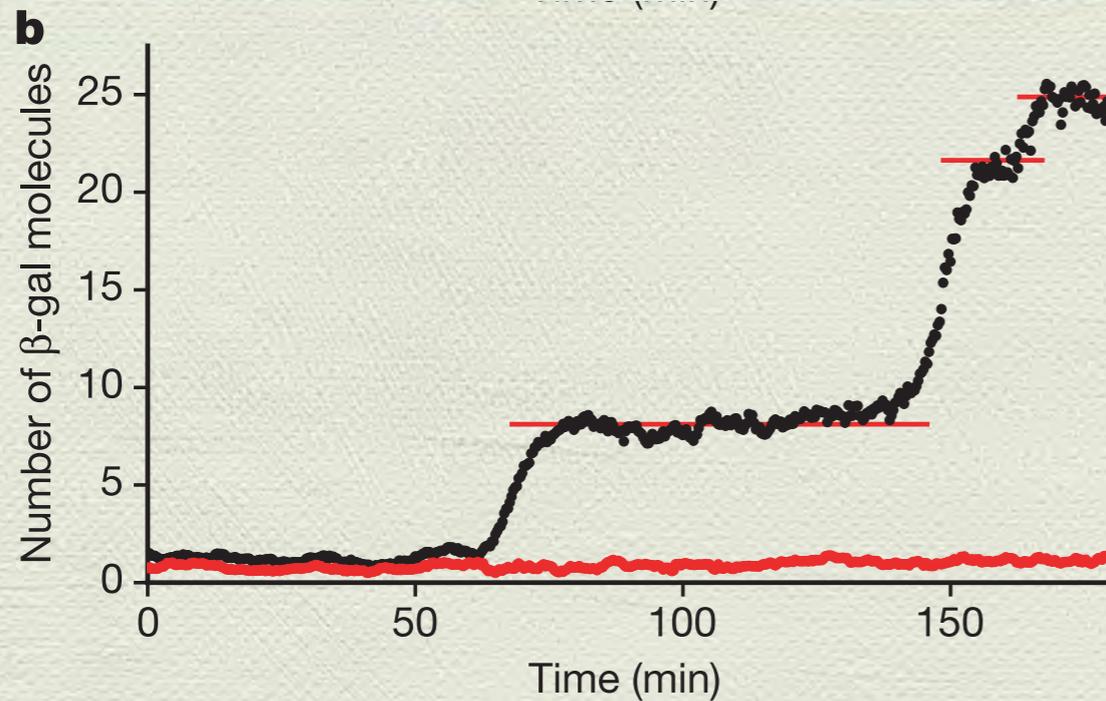
Science 304, 1811-1814 (2004)

two genes (cfp, -green; yfp, -red) controlled by identical promoters, integrated at the same locus on homologous chromosomes.

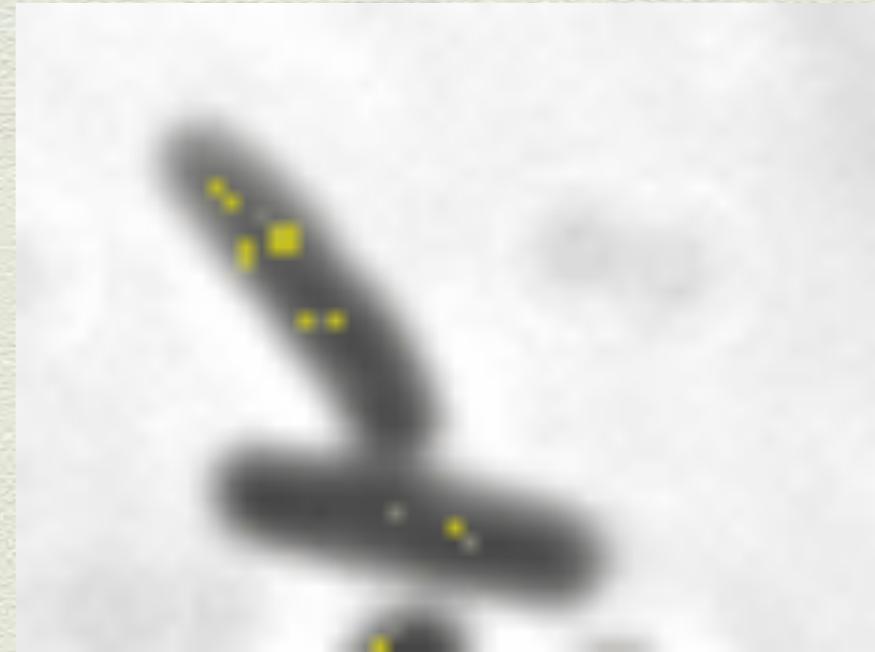
Today's talk includes

1. Theoretical Development for fluctuation and dynamics in a cell.
2. An account for gene expression noises.
-for computer simulation.
3. An application project in *C. elegans* development.

Genes express in bursts. The production is noisy.



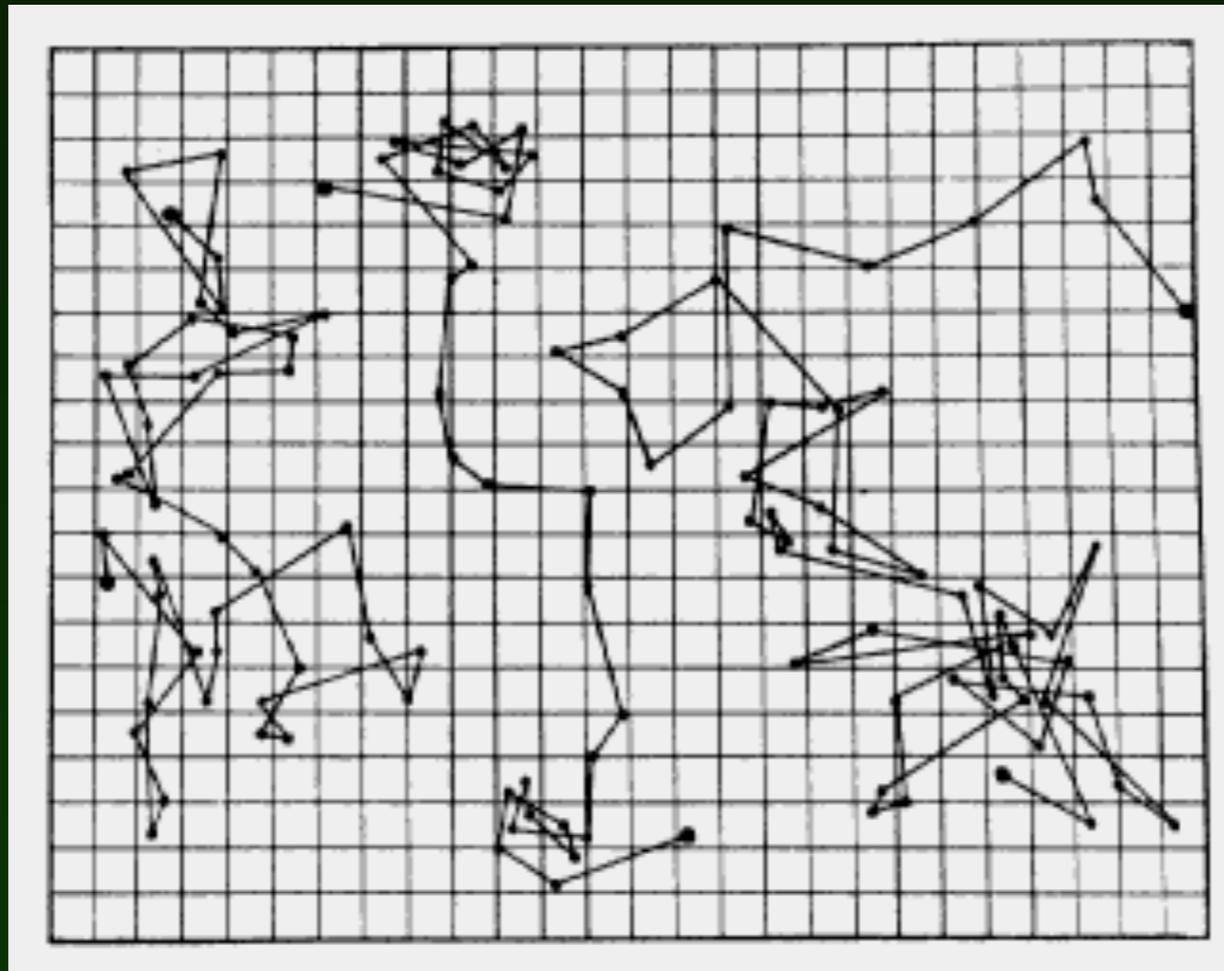
L. Cai, N. Friedman and X. S. Xie
Nature 440, 358-362 (2006)



P. Choi, L. Cai, K. Frieda and X. S. Xie
Science 322, 442-446 (2008)

Brownian motion

Coordinate
of the
particle



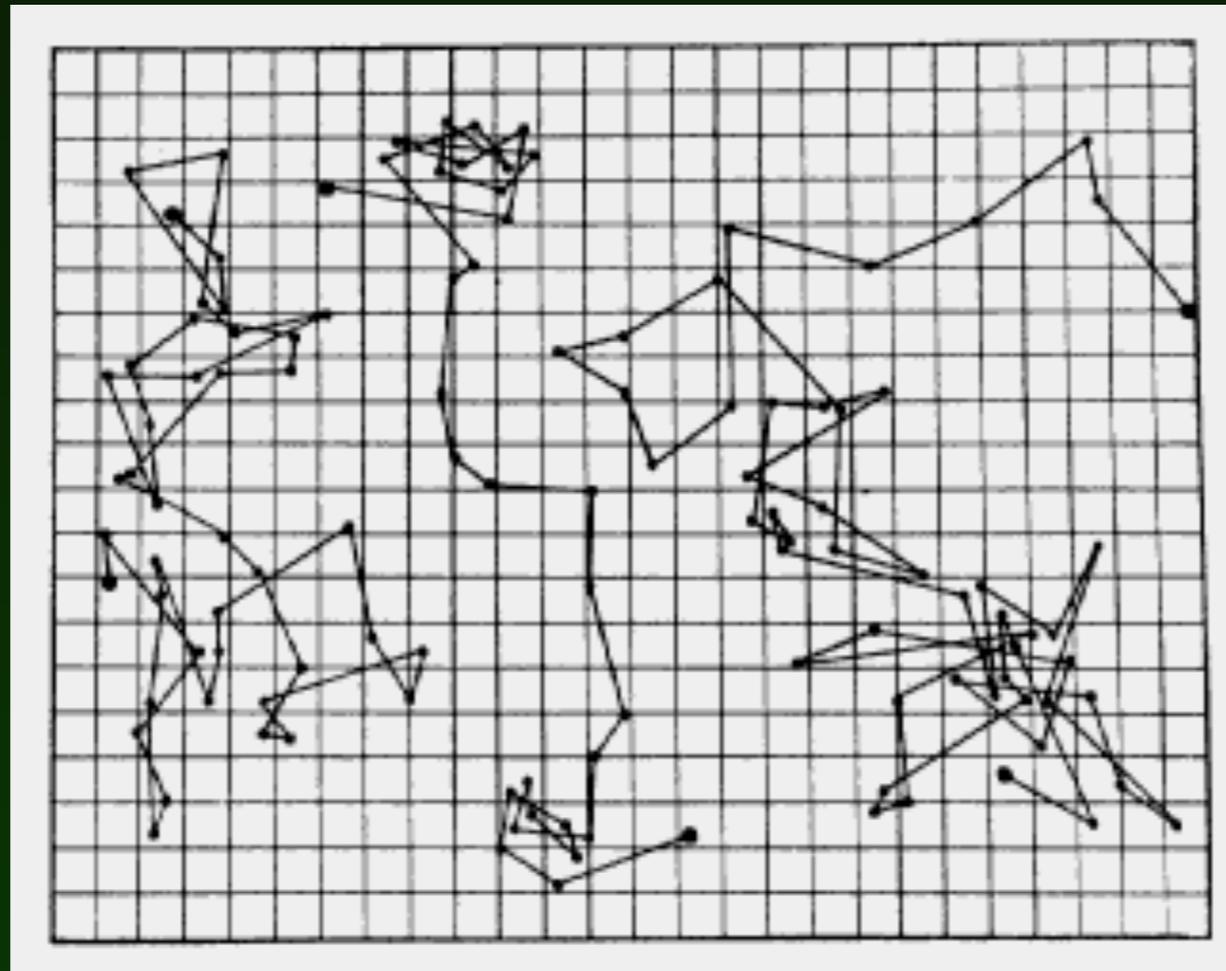
- Each step is:
 - A random "kick" from the solvent

Coordinate of the particle

Brownian motion vs. a noisy cell

Coordinate
of the
particle

Number
of
molecules
(B)



Coordinate of the particle
Number of molecules (A)

- Each step is:
 - A random "kick" from the solvent
 - ◆ A chemical reaction

Measurement of cells' response in an oscillatory perturbation

REPORTS

The Frequency Dependence of Osmo-Adaptation in *Saccharomyces cerevisiae*

Jerome T. Mettetal,¹ Dale Muzzey,^{1,2} Carlos Gómez-Uribe,^{1,3} Alexander van Oudenaarden^{1*}

Metabolic gene regulation in a dynamically changing environment

Matthew R. Bennett^{1,2*}, Wyming Lee Pang^{1,2†}, Natalie A. Ostroff¹, Bridget L. Baumgartner¹, Sujata Nayak¹, Lev S. Tsimring¹ & Jeff Hasty^{1,2}

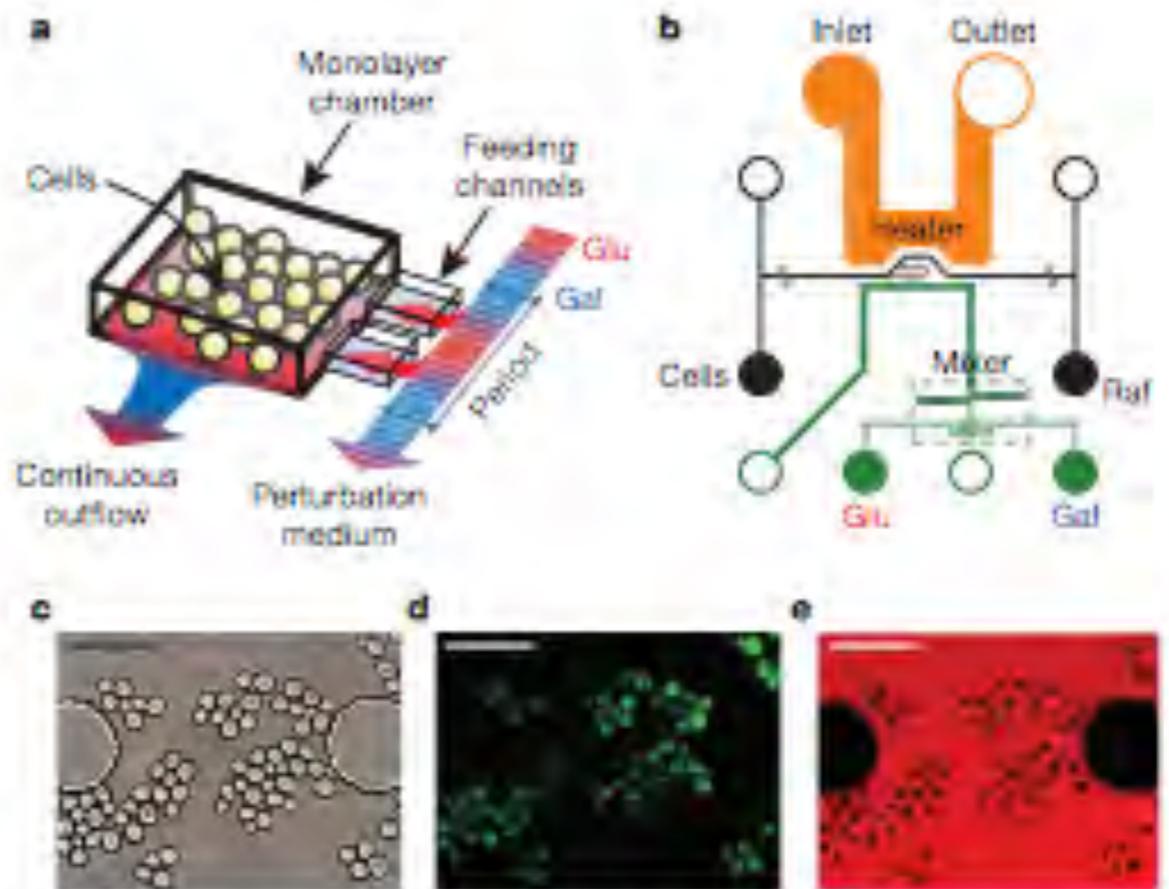
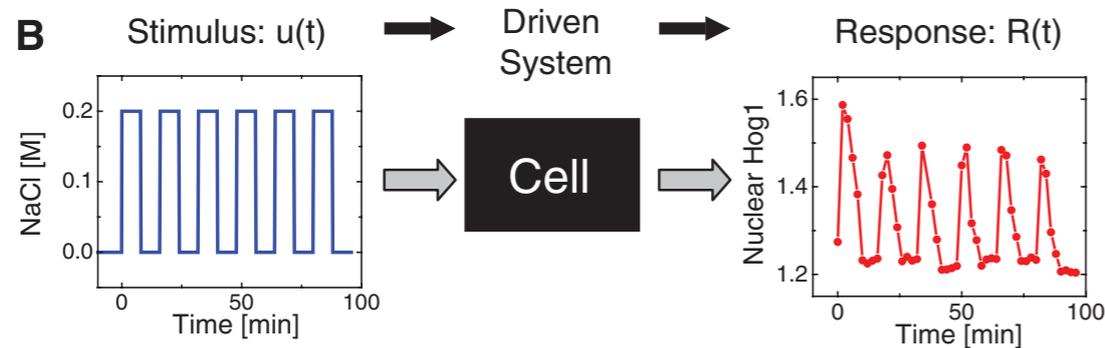
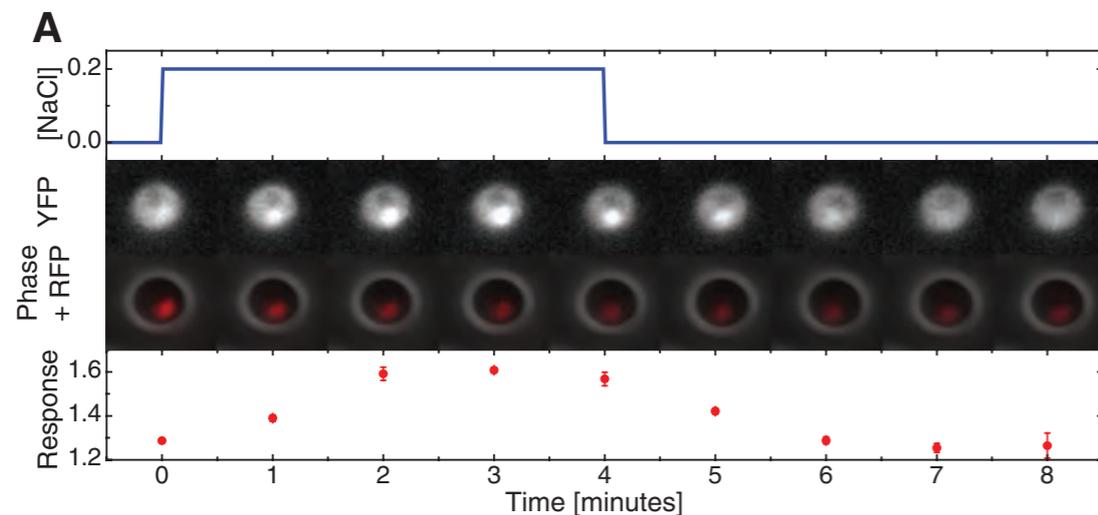
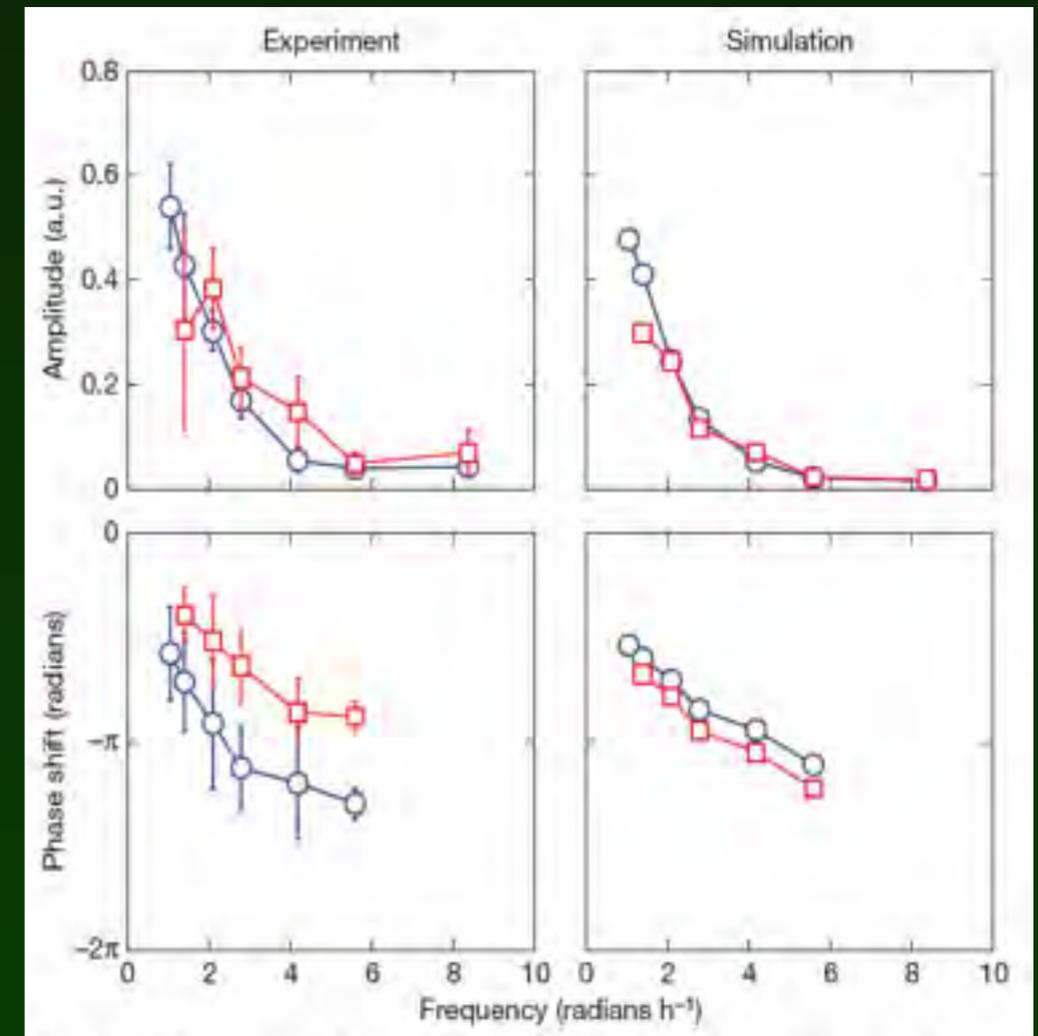
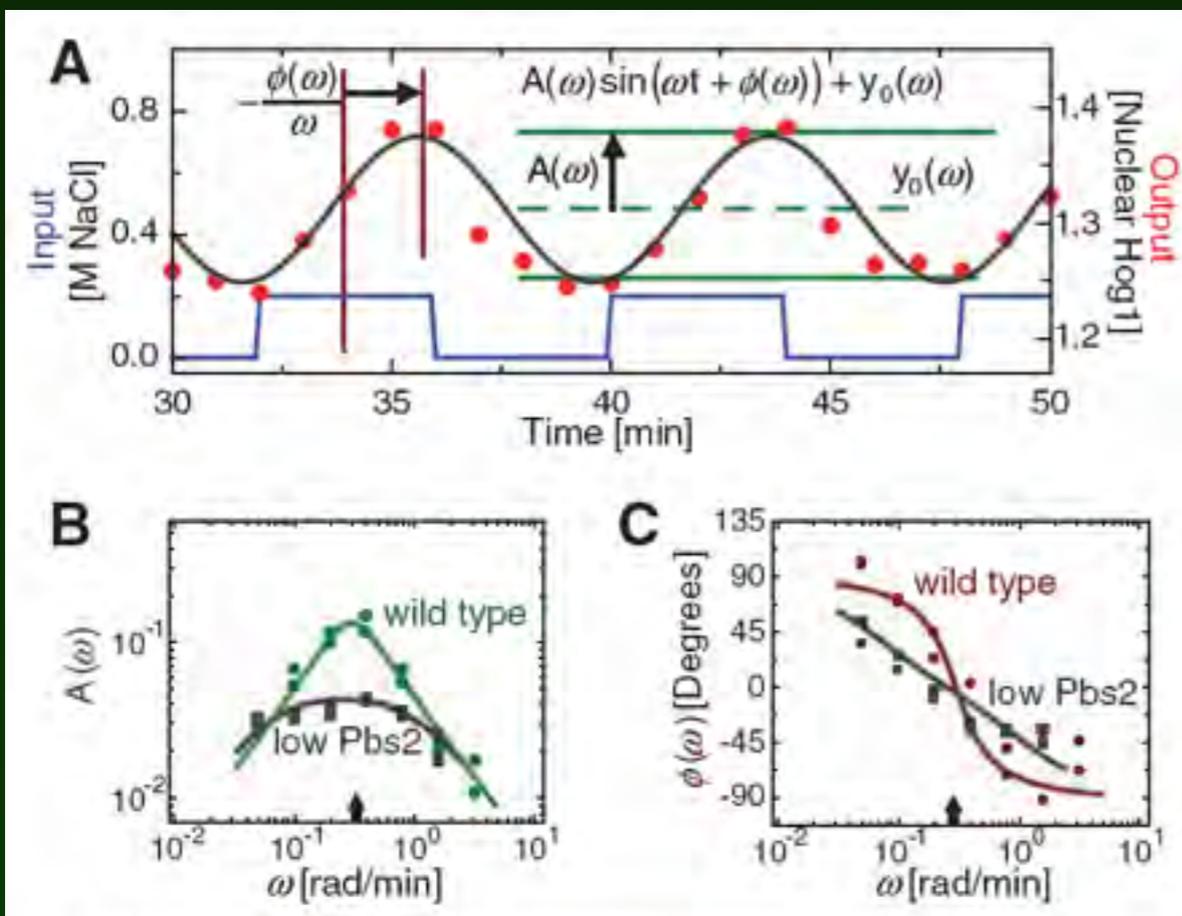
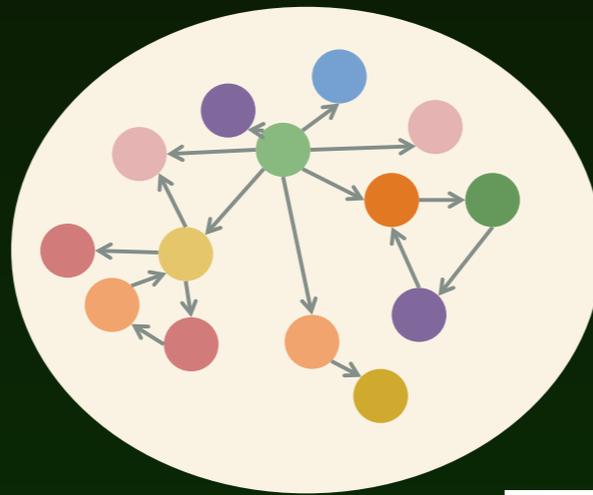


Figure 1 | Design and implementation of the microfluidic platform developed for our study. a, Conceptual design of the imaging chamber. The chamber is coupled to the switch output channel by means of multiple

"Spectroscopy" for a cell?



Linear response function

Time domain

The linear response relationship is:

$$O(t) = \int_{-\infty}^t \chi(t-t')I(t')dt'$$

$I(t)$: Time-dependent perturbation

$O(t)$: observed output

Eg. $I(t)$ = electric field

$O(t)$ = polarization in a material

$\chi(t)$ = electric susceptibility,
or dielectric constant.

Frequency domain

The linear response relationship is:

$$O(\omega) = \chi(\omega)I(\omega)$$

$I(\omega)$: Freq.-dependent perturbation

$O(\omega)$: observed output

Eg. $I(\omega)$ = electric field

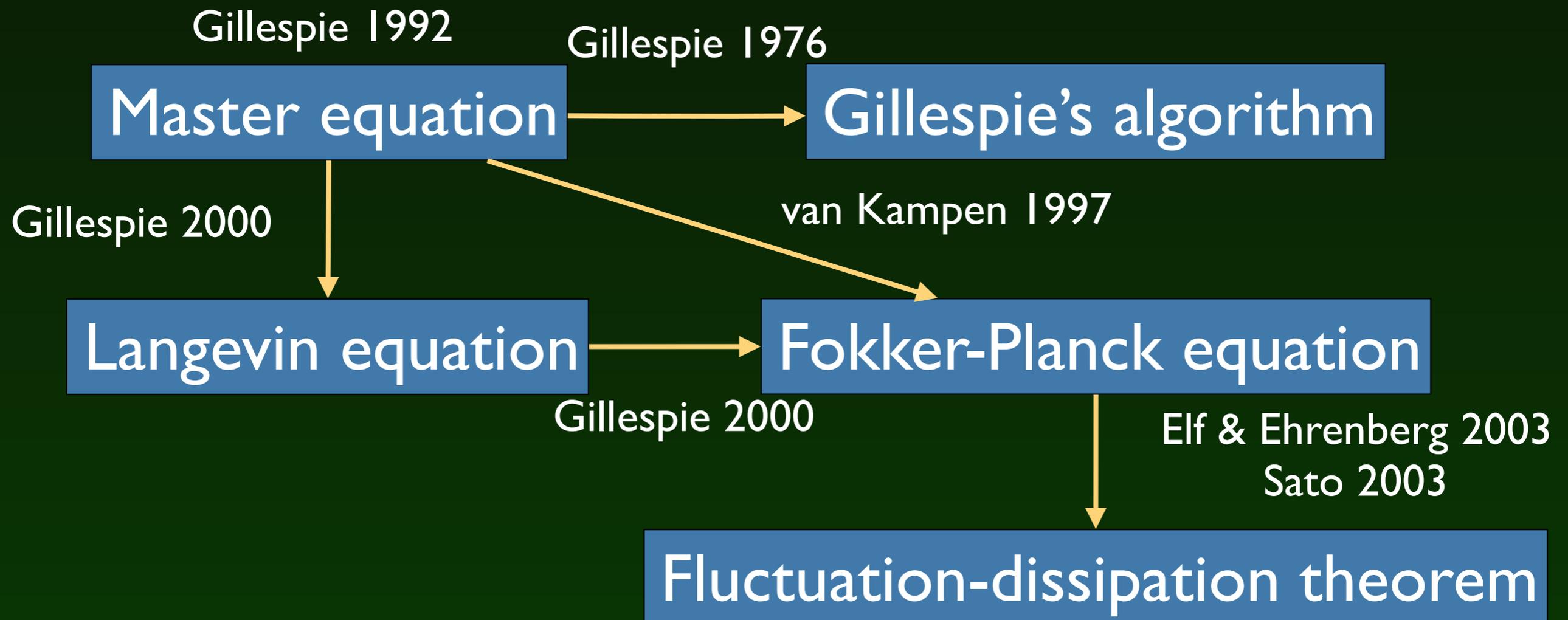
$O(\omega)$ = polarization in a material

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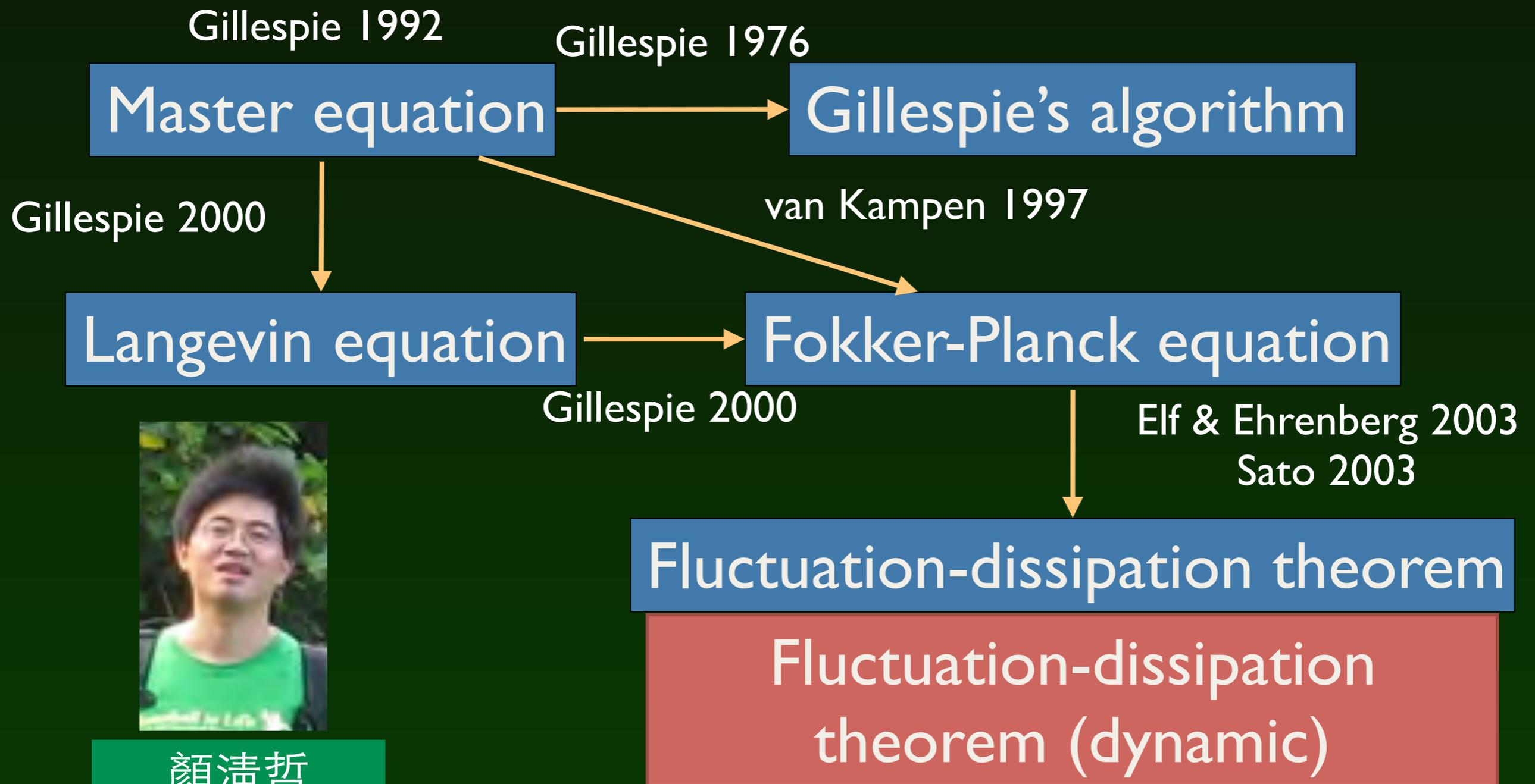
Linear Response theory in cells?

Possibility of Fluctuation-
Dissipation theorem?

Is it possible to adapt the previously developed theories/equations (in mechanics)?

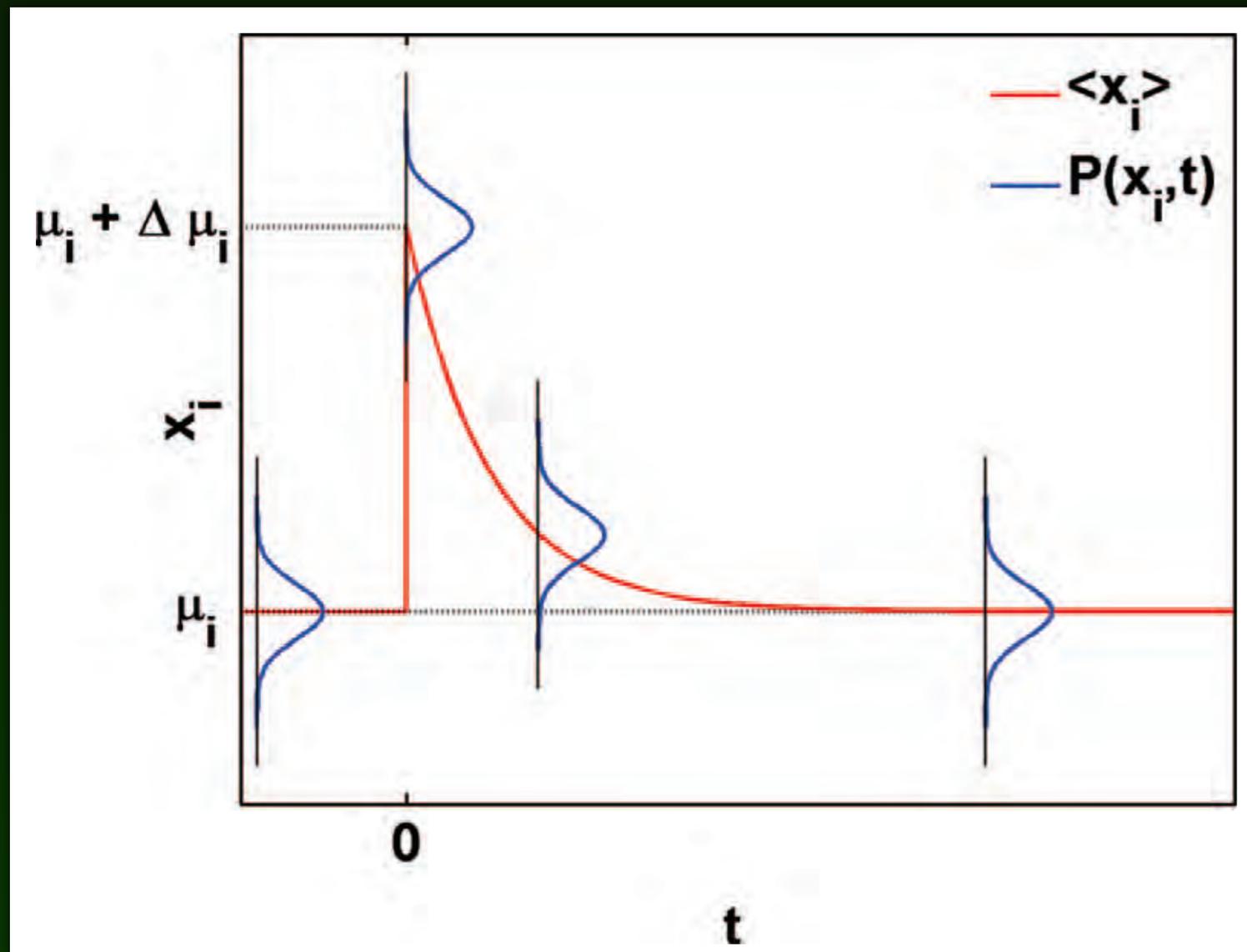


"Dynamic" version of Fluctuation-Dissipation Theorem exists



顏清哲
Sanders Yan

“Dissipation”: back to steady state



Can be simulated directly.
Can we “predict” it with FDT (i.e. with correlation function)?

Derivation for FDT

- Suppose a perturbation is applied at $t=0$: (I.e. increase or decrease the particle number in a reaction system) $\Delta\mu$
- Gaussian probability distribution assumed.
- The time-dependent change in the averaged $x(t)$ is:
$$\langle \Delta x_j(t) \rangle = \langle x_j(t) - \mu_j \rangle = \int (x_j - \mu_j) P(\mathbf{x}, t) d\mathbf{x}$$

Dynamic Fluctuation-Dissipation Theorem for biochemical kinetics

$$\bar{x}(t) - \mu_0 = \frac{\Delta\mu}{\Sigma^2} \langle \delta x(0) \delta x(t) \rangle$$

where $x(t)$ = the particle number for the observed species

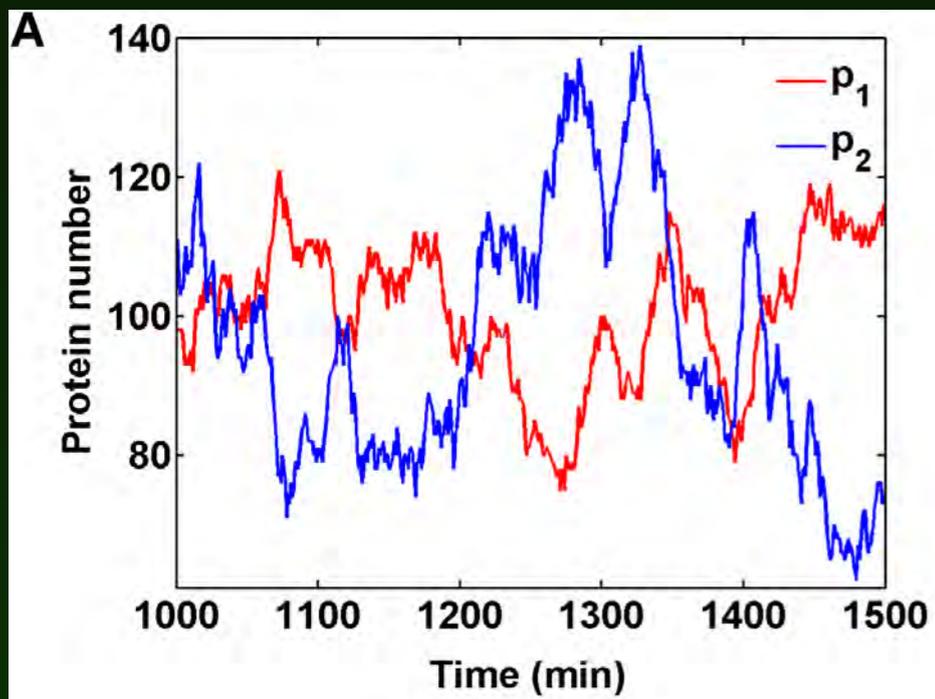
μ_0 = equilibrium average for x

Σ^2 = equilibrium variance for x

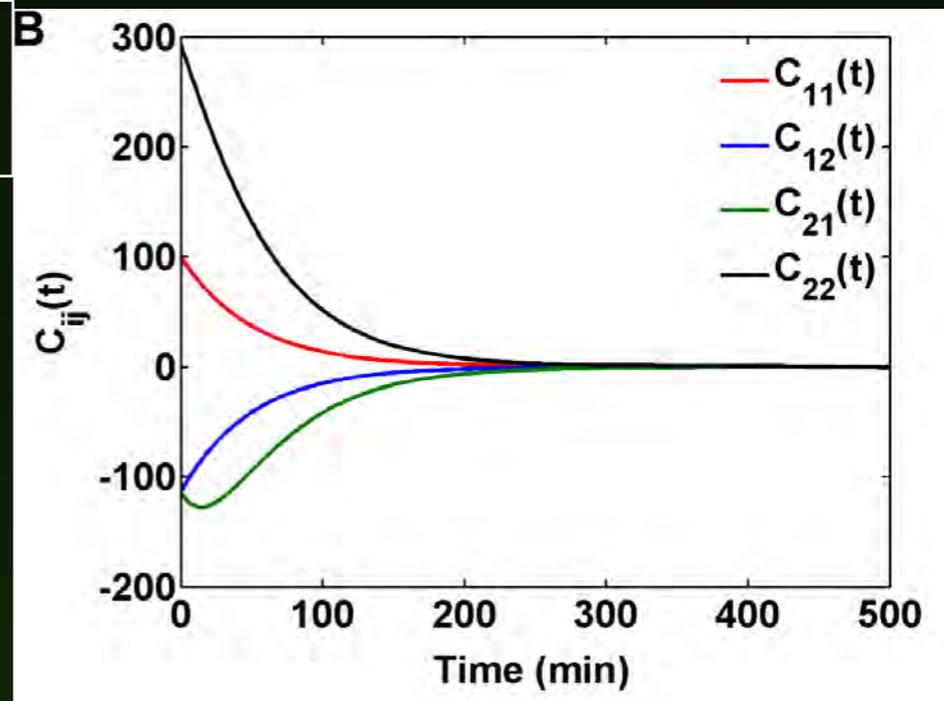
$\delta x(t)$ = instantaneous fluctuation for x

- LHS: How $x(t)$ comes back to the equilibrium, after a perturbation. A non-equilibrium quantity.
- RHS: The correlation function is the characteristic of fluctuation in $x(t)$. An equilibrium quantity.
- A linear response theory is provided.
- Can be generalized to multiple component systems.

Protein 1 (p_1) \rightarrow Protein 2 (p_2)

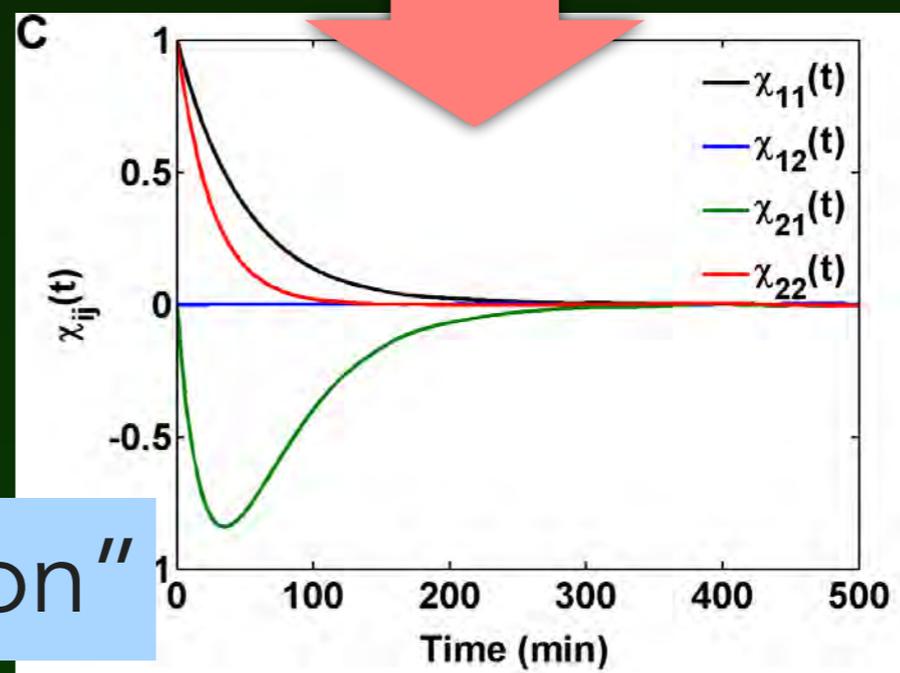


Calculate



Correlation Functions

FDT



One stochastic trajectory obtained in the steady state.

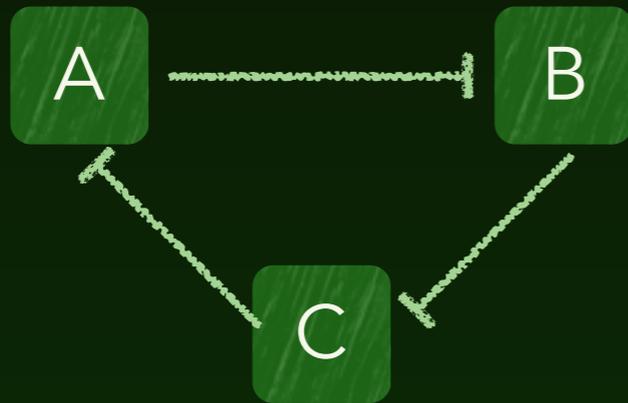
"Fluctuation"

"Dissipation"

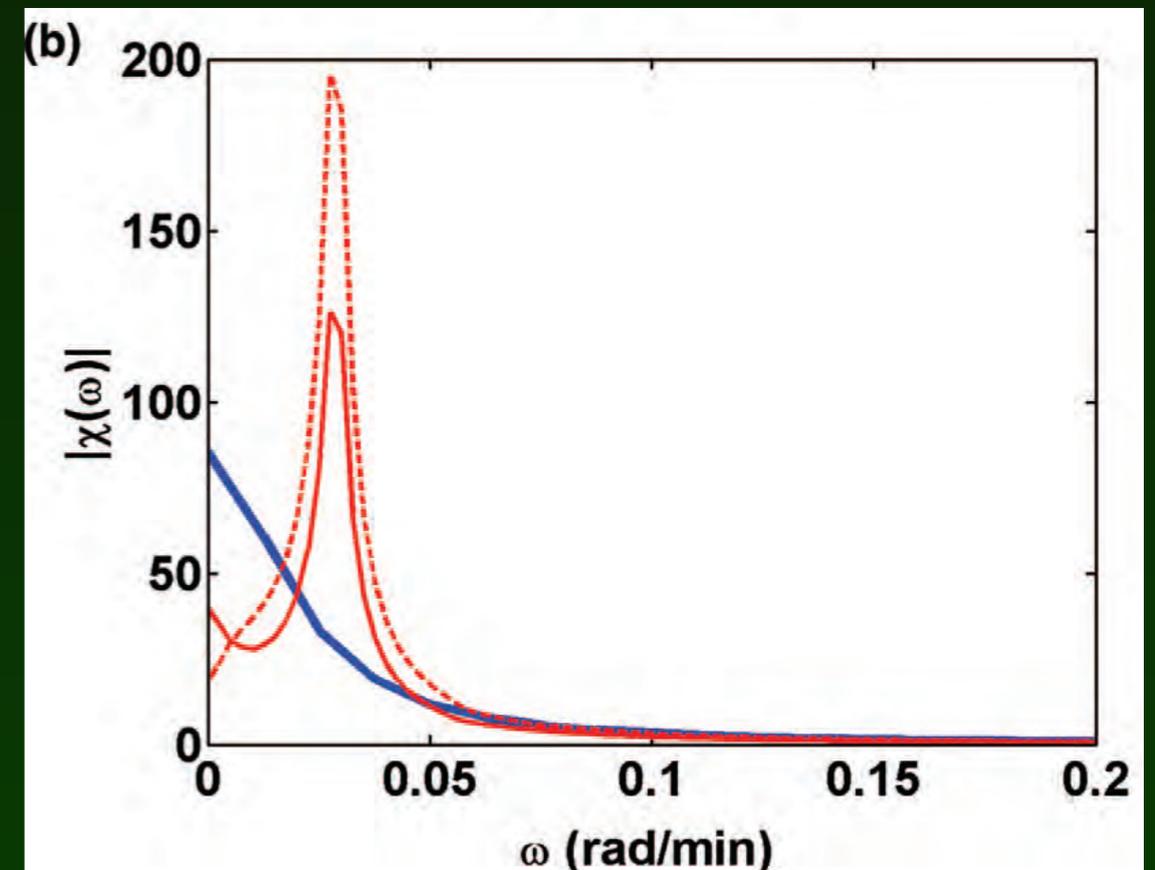
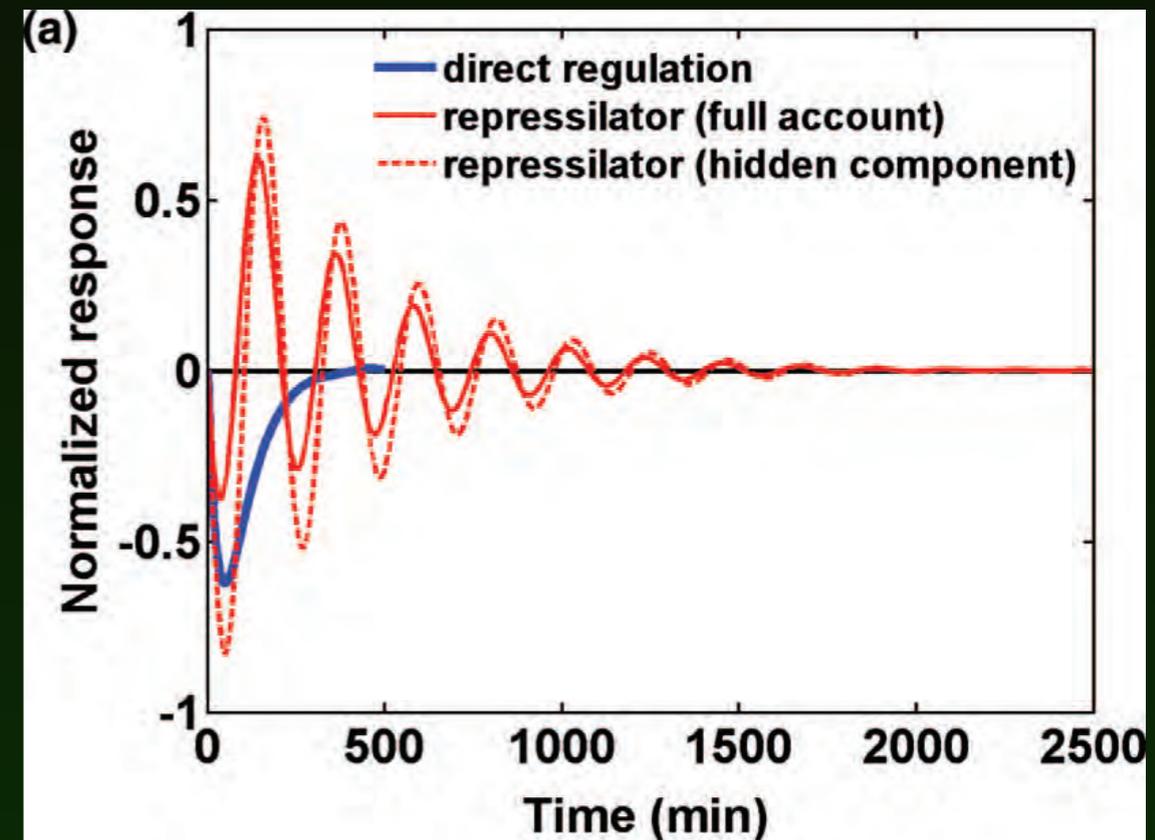
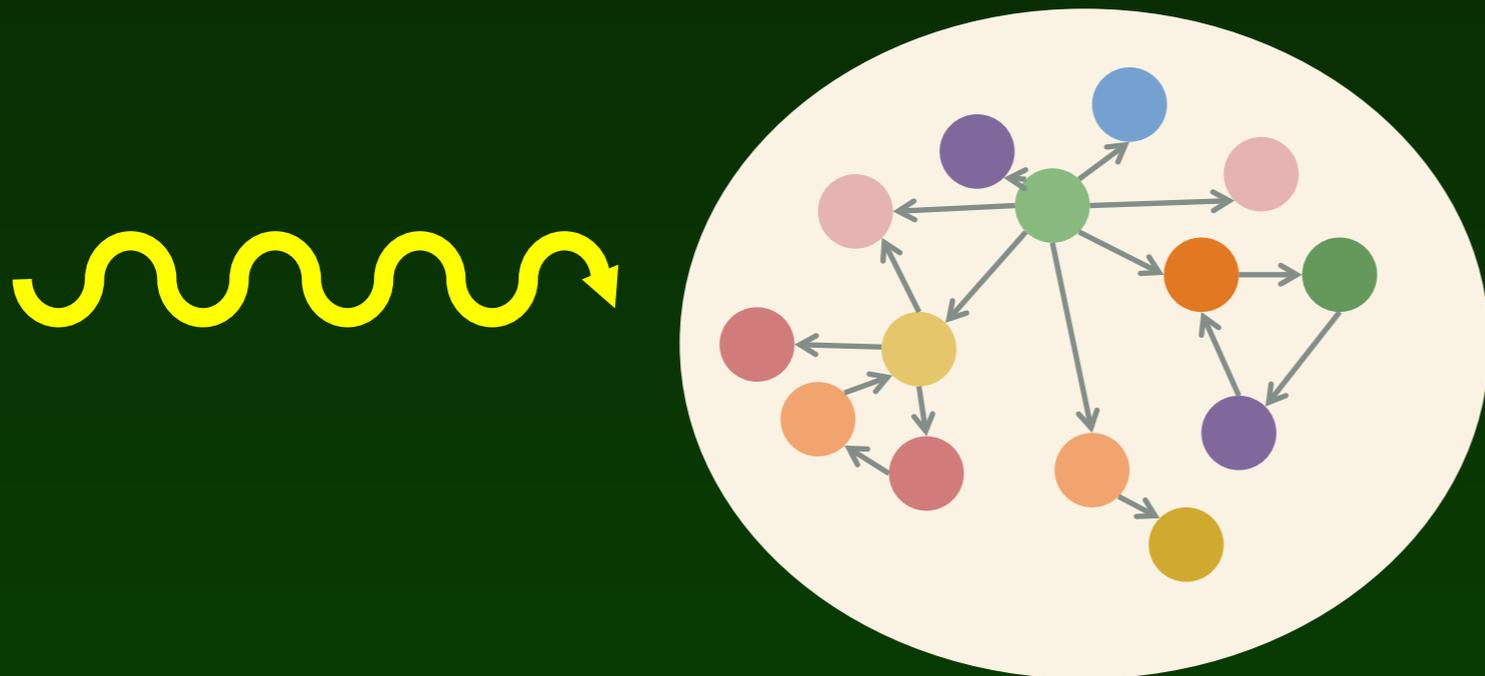
Predict the response after a perturbation.

"Spectroscopy" for a cell?

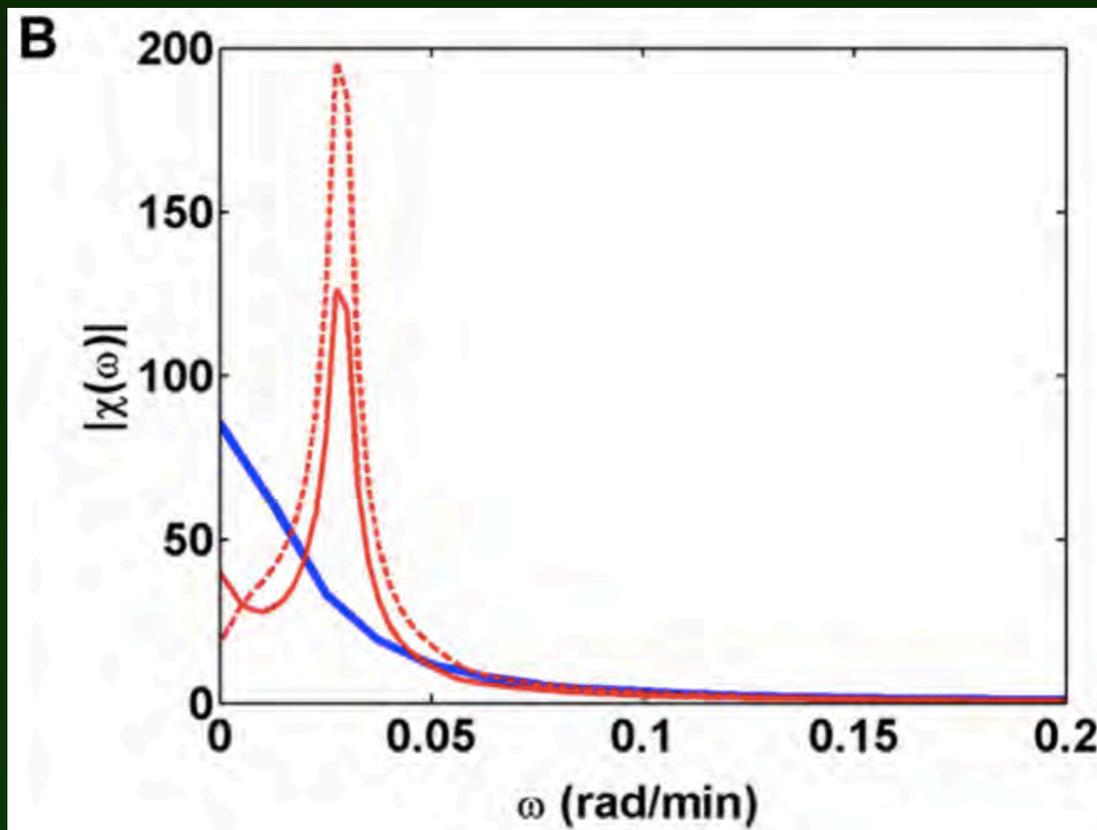
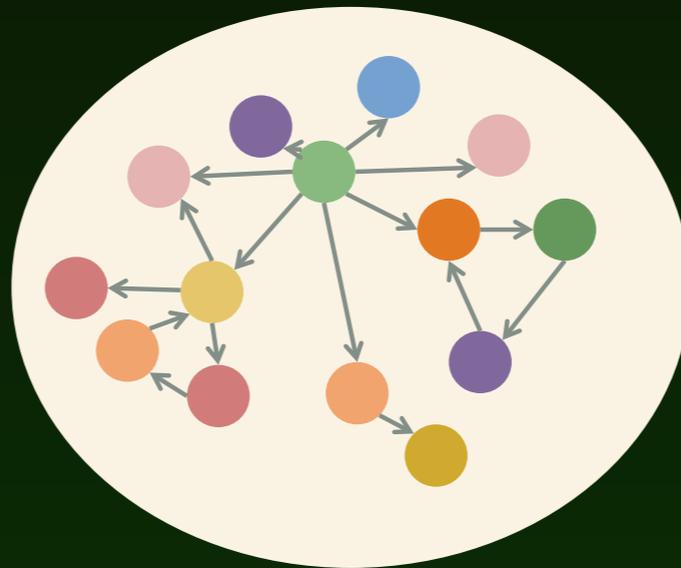
Repressillator:
a model with
intrinsic
oscillation



- A "resonance" is seen with periodic perturbation.
- Probing oscillators in a cell.



"Spectroscopy" for a cell?

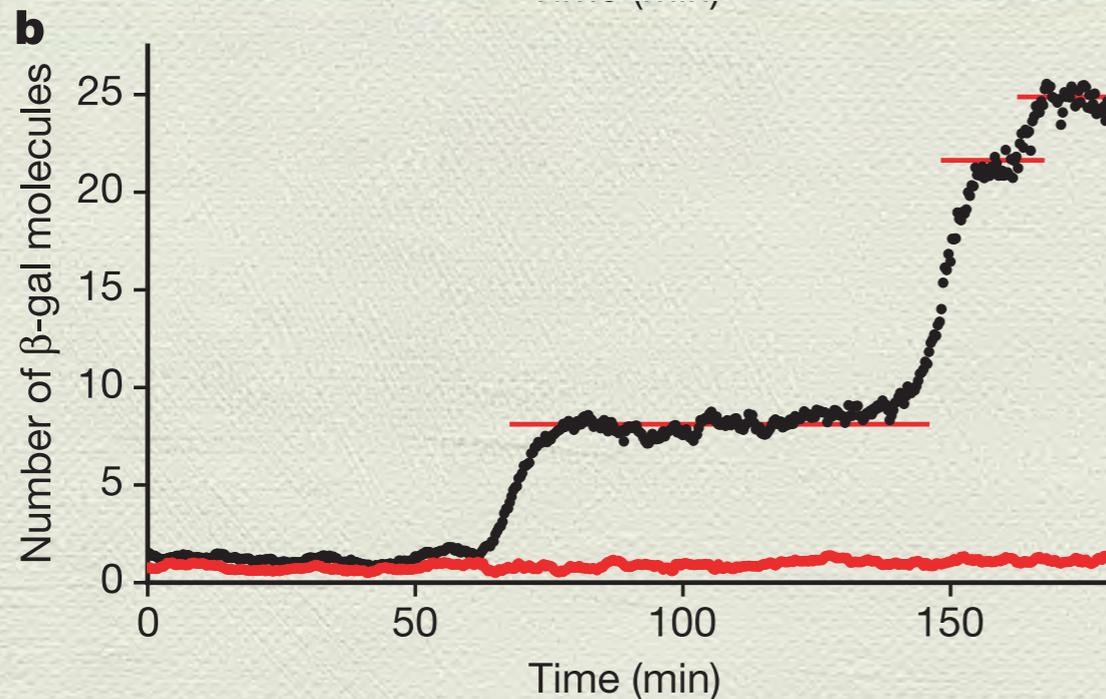


- Time scale separation?
- Not quite. Observable oscillation are mostly in 10^0 - 10^3 minutes.
- Relaxation (and dilution due to cell div.), also in 10^1 - 10^3 minutes.

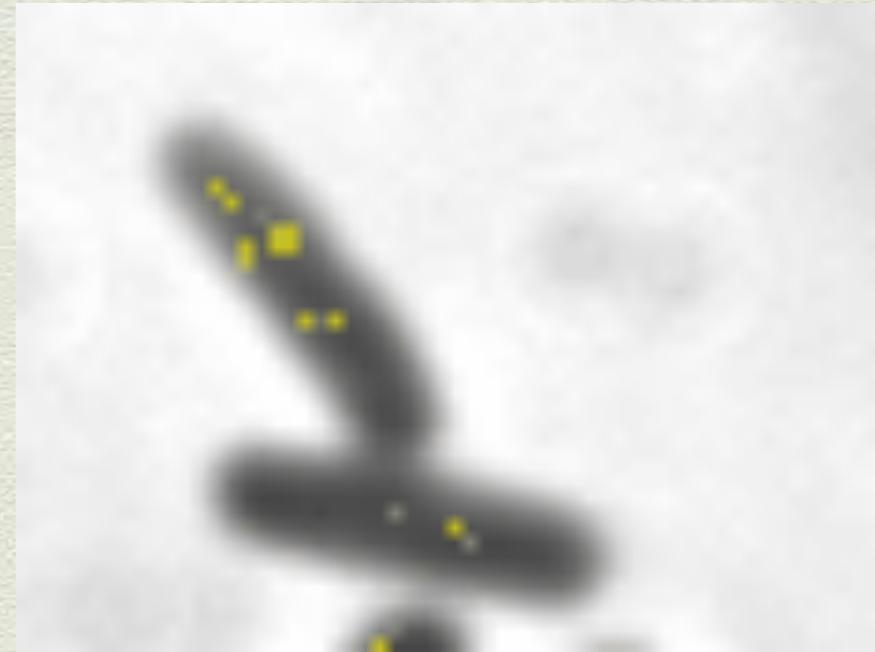
Summary

- FDT — it is possible use correlation functions to compose the response function.
- To predict response from fluctuation, linear response is necessary.
- There is a limited range for linear response.
- (data not shown) It is possible to construct the response function for missing component situation.
- Limited data sampling is not a big problem either.

Genes express in bursts. The production is noisy.

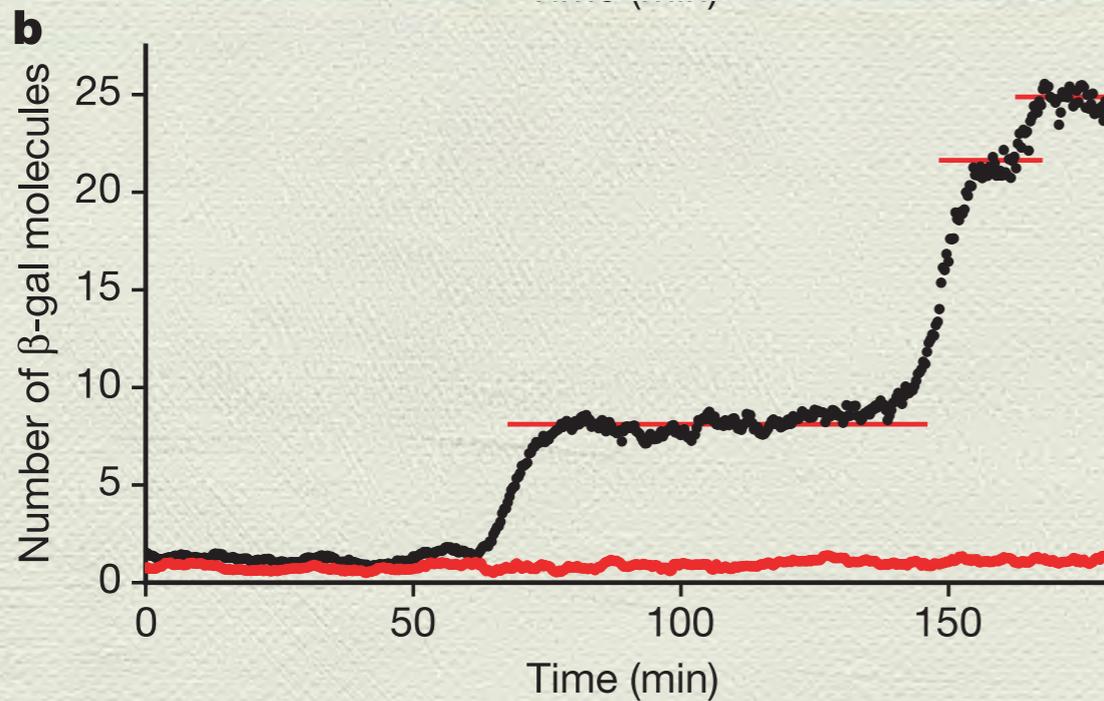


L. Cai, N. Friedman and X. S. Xie
Nature 440, 358-362 (2006)

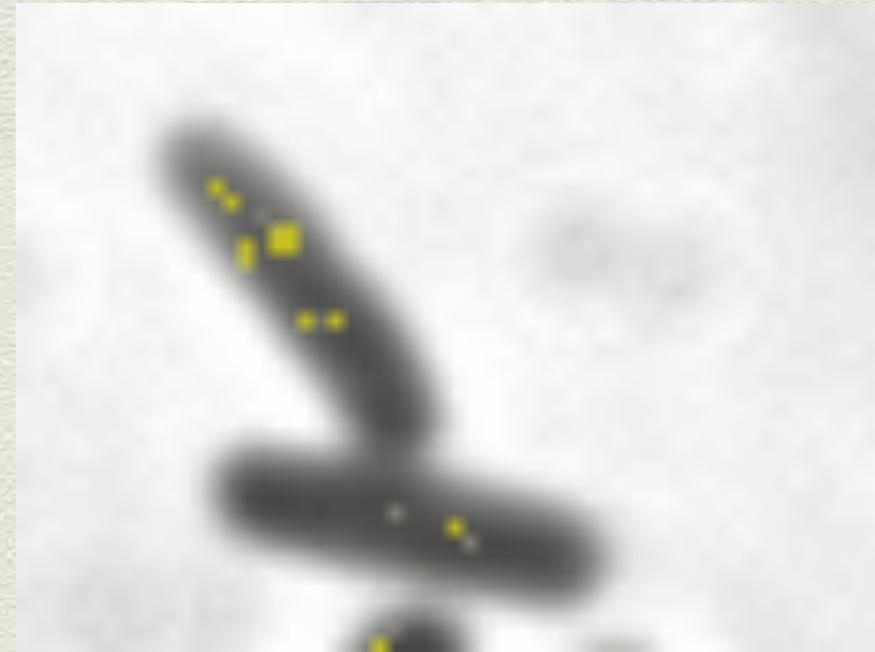


P. Choi, L. Cai, K. Frieda and X. S. Xie
Science 322, 442-446 (2008)

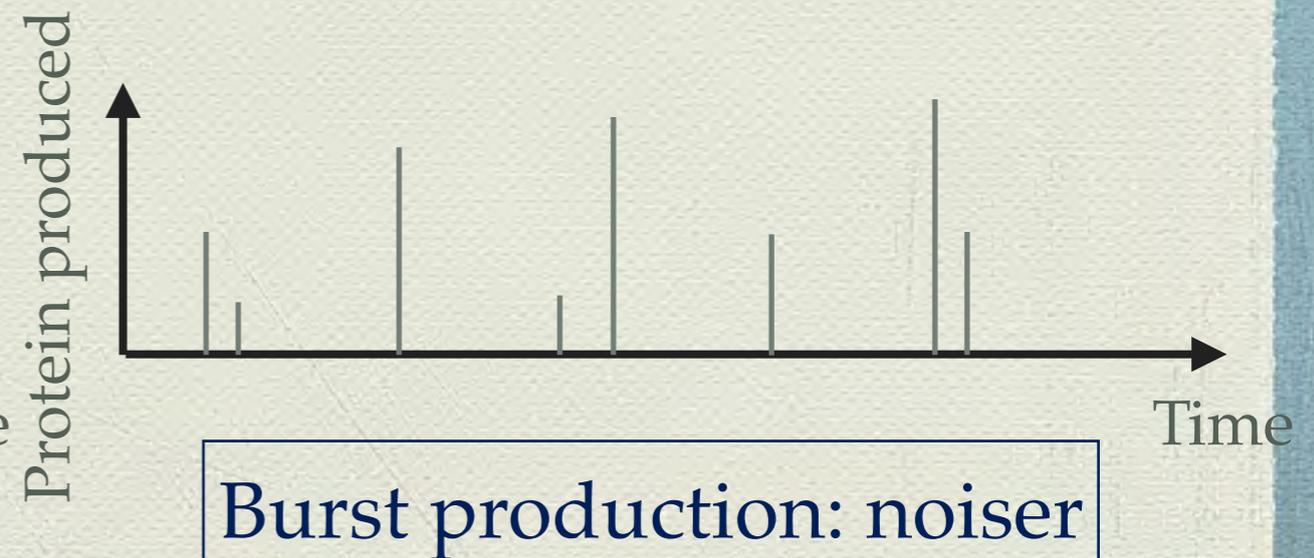
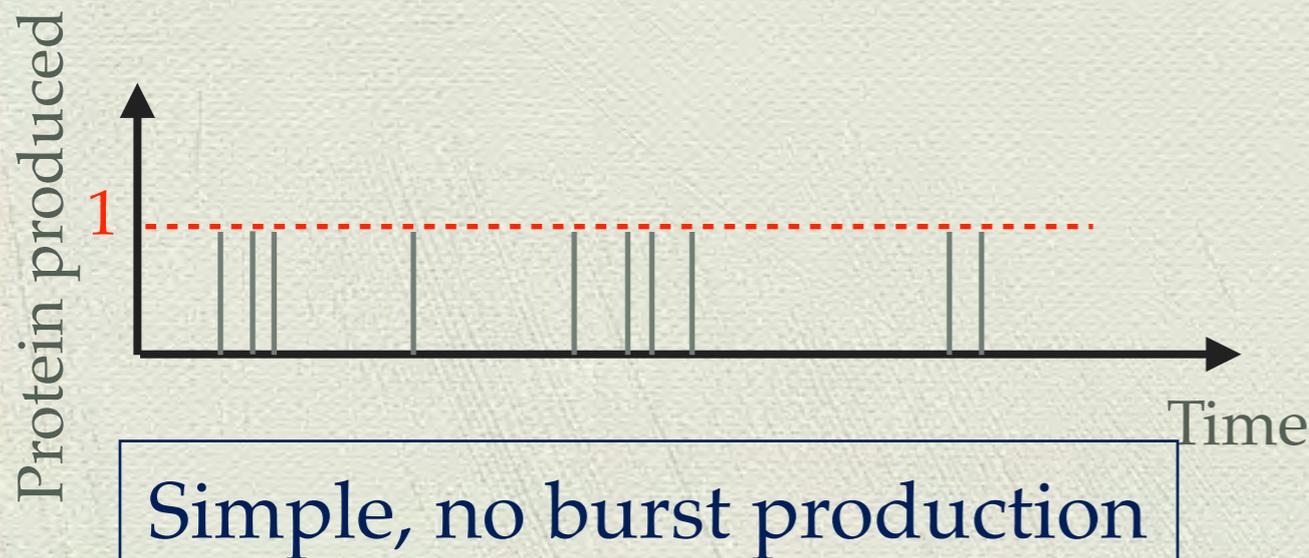
Genes express in bursts. The production is noisy.



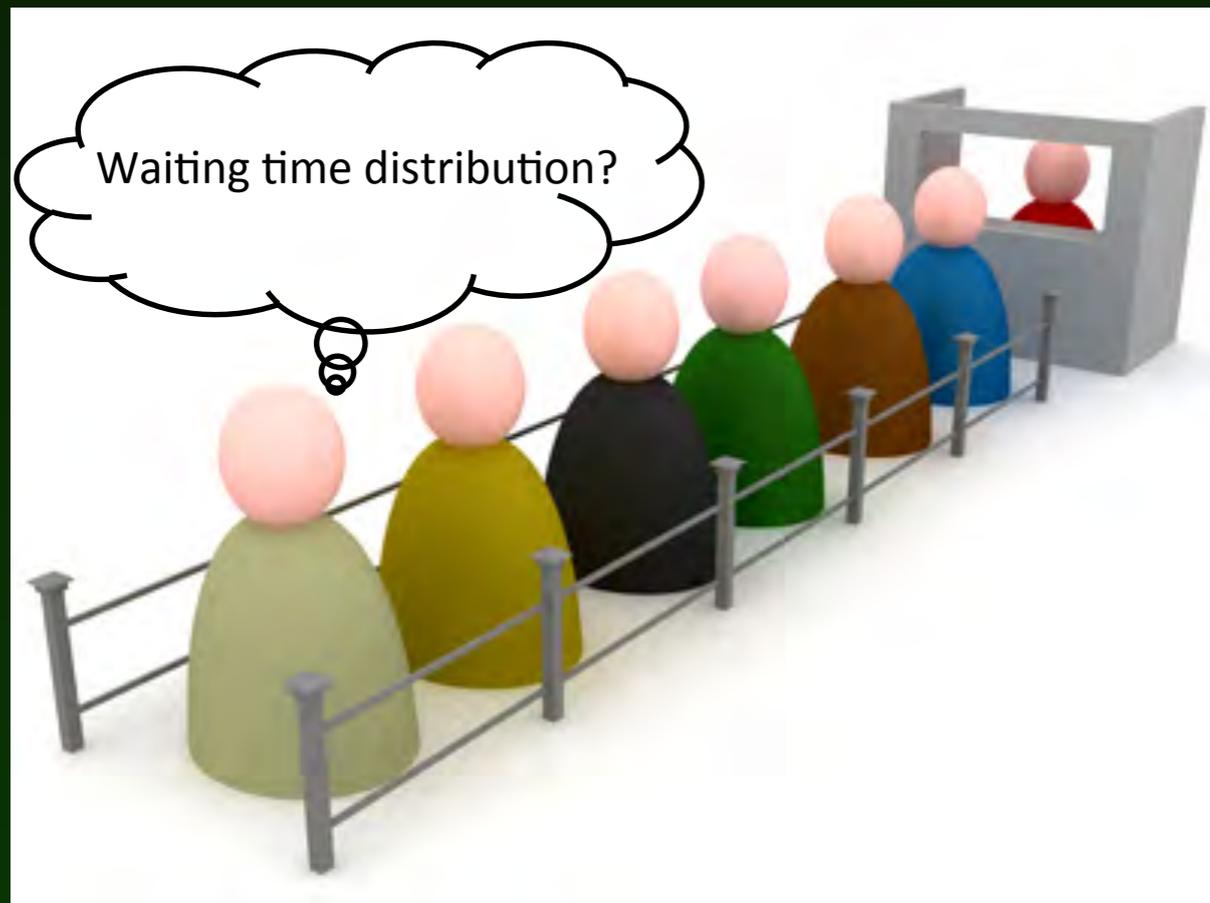
L. Cai, N. Friedman and X. S. Xie
Nature 440, 358-362 (2006)



P. Choi, L. Cai, K. Frieda and X. S. Xie Science
322, 442-446 (2008)



The distribution of protein expressed



- Number of people in front of you ~ Poisson distribution. (Number of bursts)
- Time each person spend ~ exponential distribution (Sunney Xie's result: burst size distribution).
- Total waiting time (total protein produced) distribution?

Figure from:

<http://chaaidaani.wordpress.com/2012/05/02/good-mannered-when-abroad-bad-mannered-in-pakistan/>

Langevin's equation

Protein burst model

- $dm/dt = k_m - \gamma_m m$
- $dp/dt = k_p m - \gamma_p p$

A Gaussian
random number

$$p(t + \tau) = p(t) + \left[k_p m \tau + (k_p m \tau)^{1/2} N(0, 1) \right] \\ - \left[\gamma_p p \tau + (\gamma_p p \tau)^{1/2} N(0, 1) \right]$$

Regular reaction channels: Poisson statistics

Langevin's equation, without having to model m (=mRNA)

Burst channel

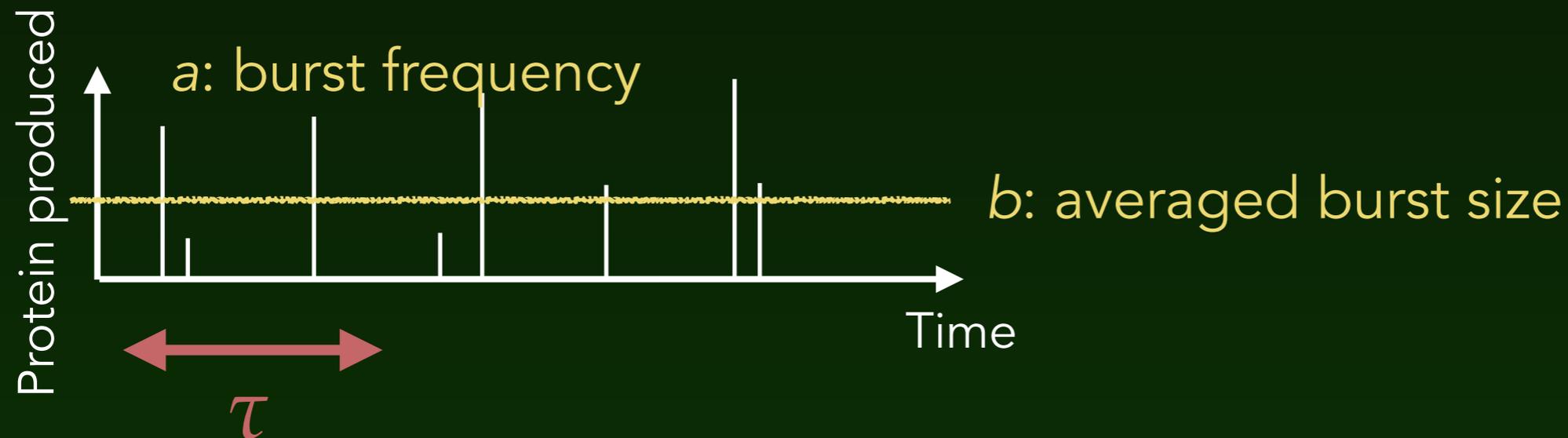
$$p(t + \tau) = p(t) + \sum_{i=1}^{n_e} x_i$$

$$\Rightarrow \mu_p + \sigma_p N(0,1)$$

$$- \left[\gamma_p p \tau + (\gamma_p p \tau)^{1/2} N(0,1) \right]$$

Non-burst, "normal" channels

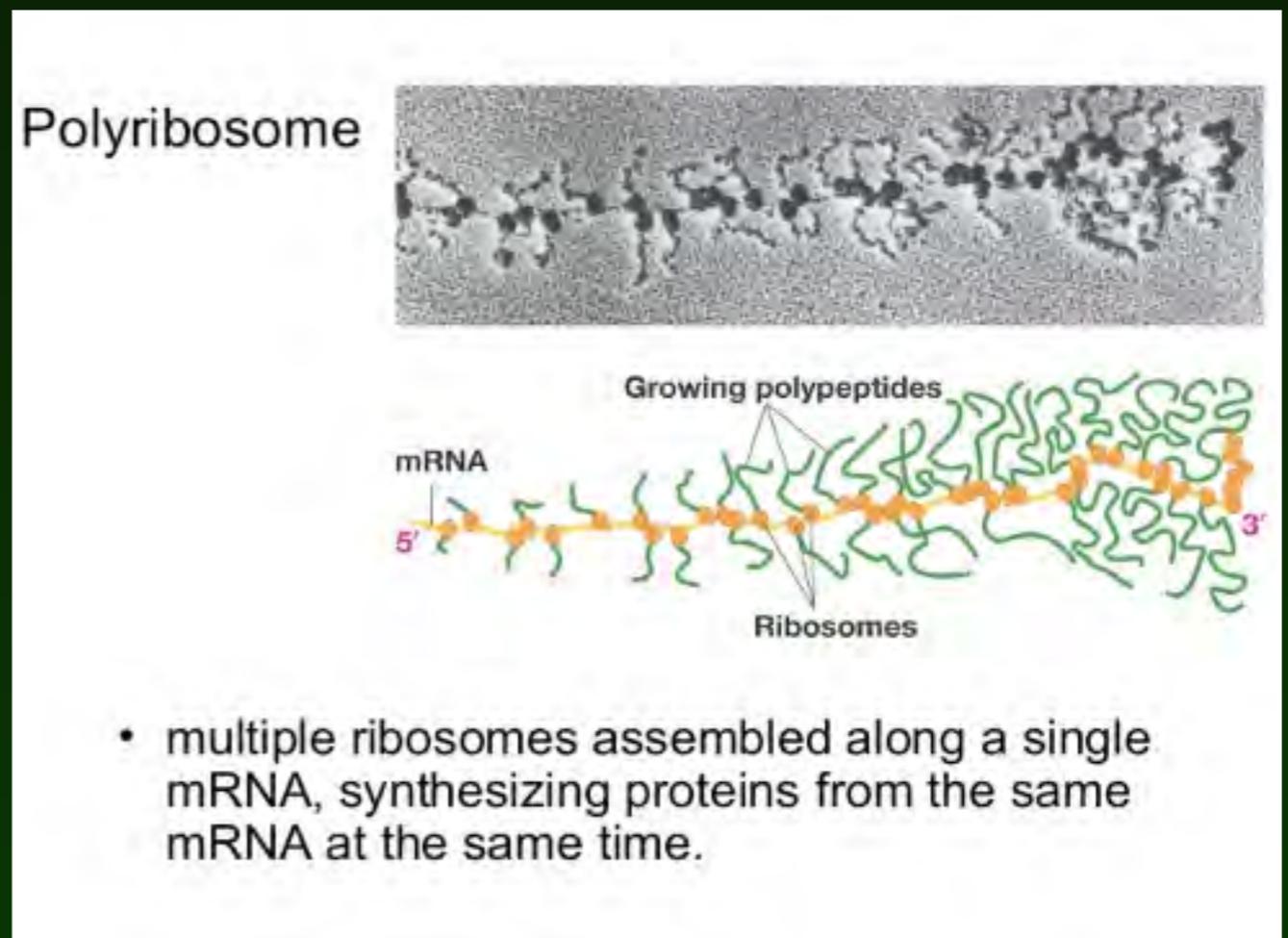
How many protein is produced in time τ ?



- in time τ , on the avg, $a\tau$ bursts.
- Total: $a\tau b$ protein on the average.
- What is the variance of this total # of protein?

Some Basics

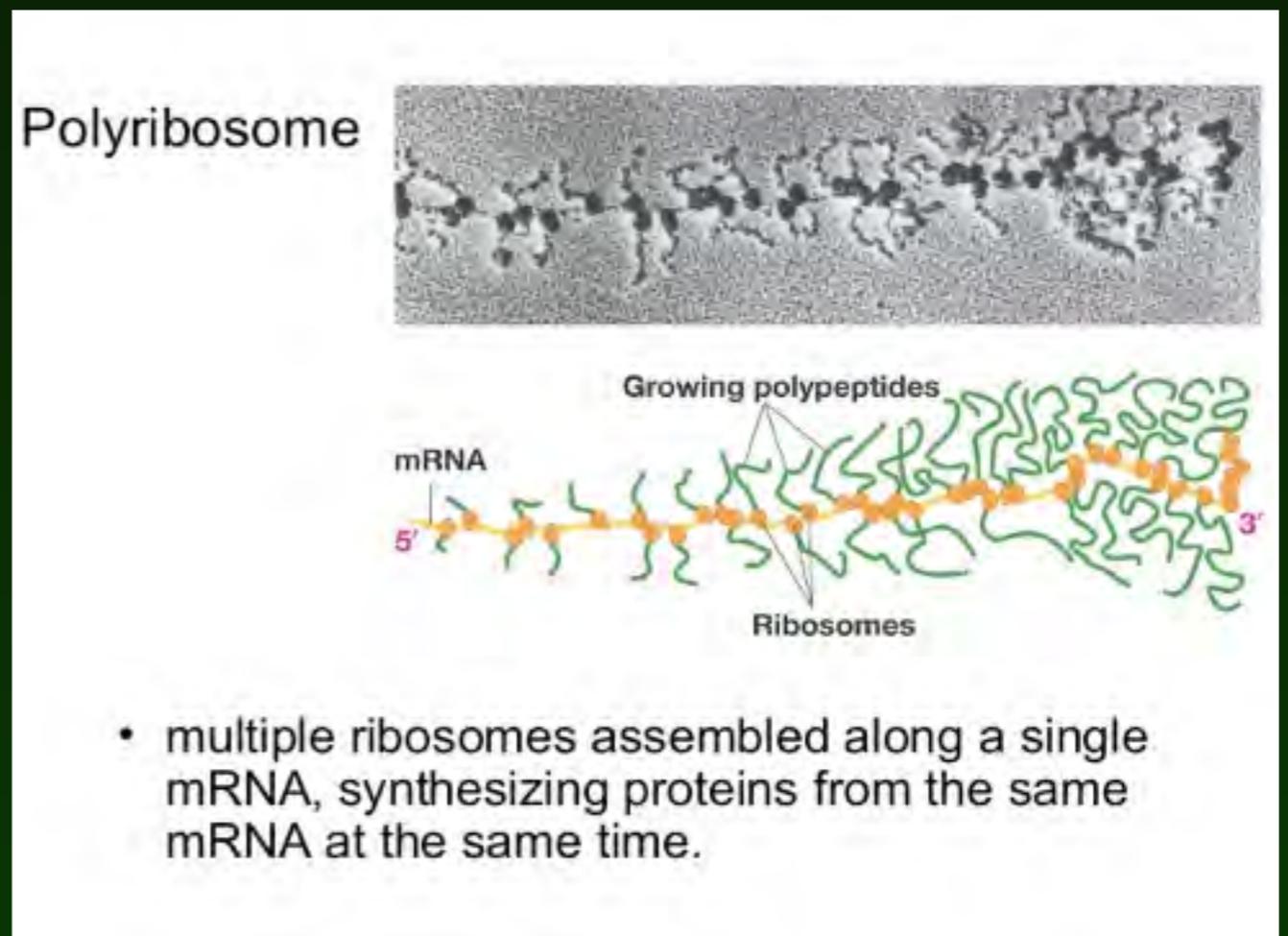
- Protein burst model
 - $dm/dt = k_m - \gamma_m m$
 - $dp/dt = k_p m - \gamma_p p$
- mRNA degrades fast:
 $\gamma_m \gg \gamma_p$
- mRNA production is slow: $k_m \ll k_p$
- 1 mRNA \approx 1 burst protein production.



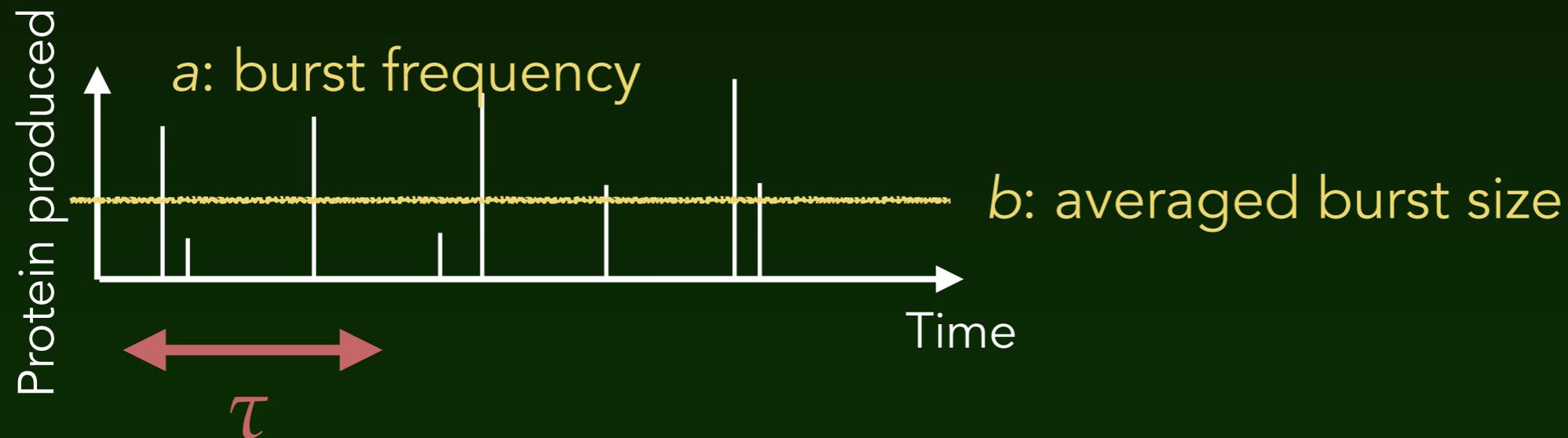
Some Basics

- Protein burst model
 - $dm/dt = k_m - \gamma_m m$
 - $dp/dt = k_p m - \gamma_p p$

- Protein production rate
= $k_p m = k_p k_m / \gamma_m$
= $k_m (k_p / \gamma_m)$
- $a = k_m$
- $b = k_p / \gamma_m$
- each mRNA produces k_p / γ_m protein. This # is exp. in distribution.



How many protein is produced in time τ ?



- in time τ , on the avg, $a\tau$ bursts.
- Total: $a\tau b$ protein on the average. $a\tau b$
- What is the variance of this total # of protein? $2a\tau b^2$

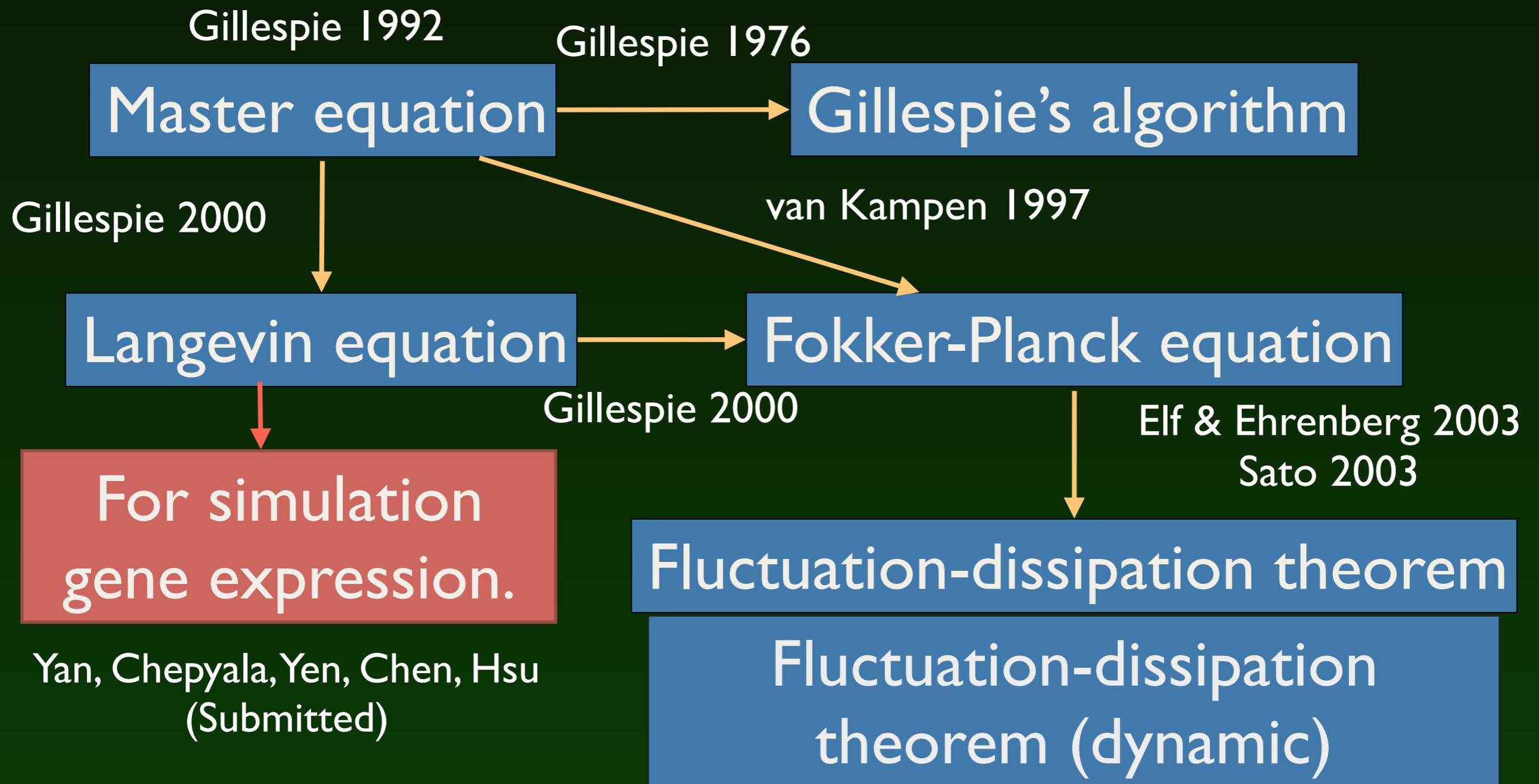
Langevin's equation can be formulated

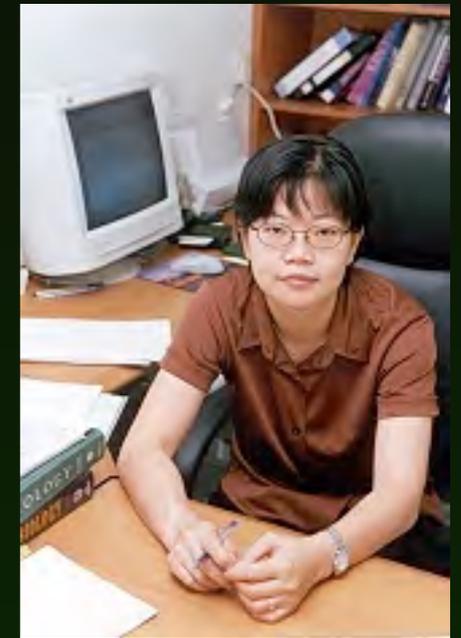
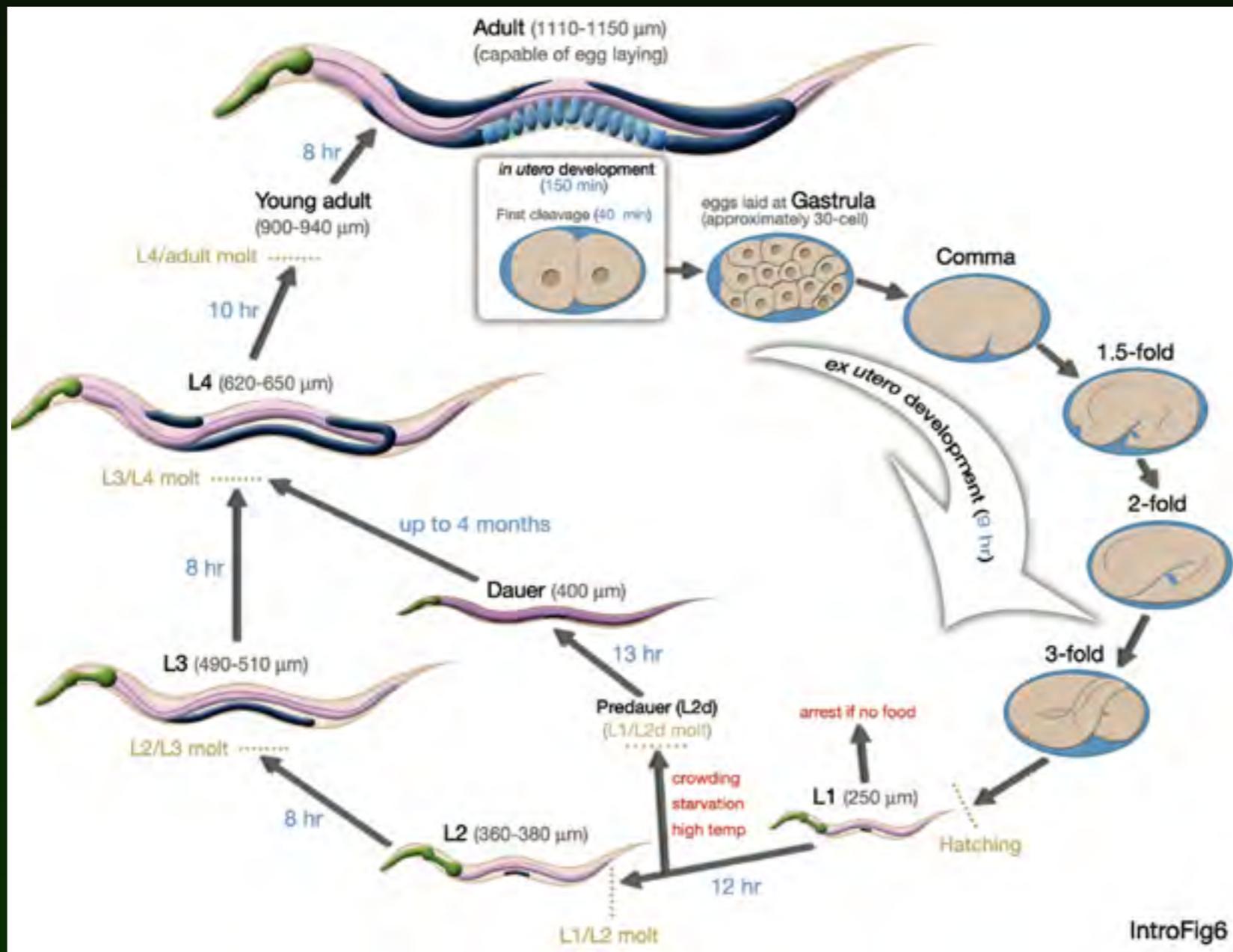
Burst channel

$$p(t + \tau) = p(t) + \left(a\tau b_p + (a\tau b_p (2b_p + 1))^{1/2} N(0, 1) \right) \\ - \left(\gamma_p p\tau + (\gamma_p p\tau)^{1/2} N(0, 1) \right),$$

Non-burst, "normal" channels

The noise of a bursting gene production is $\sqrt{2}$ times the no-burst (1 copy at a time) noise.





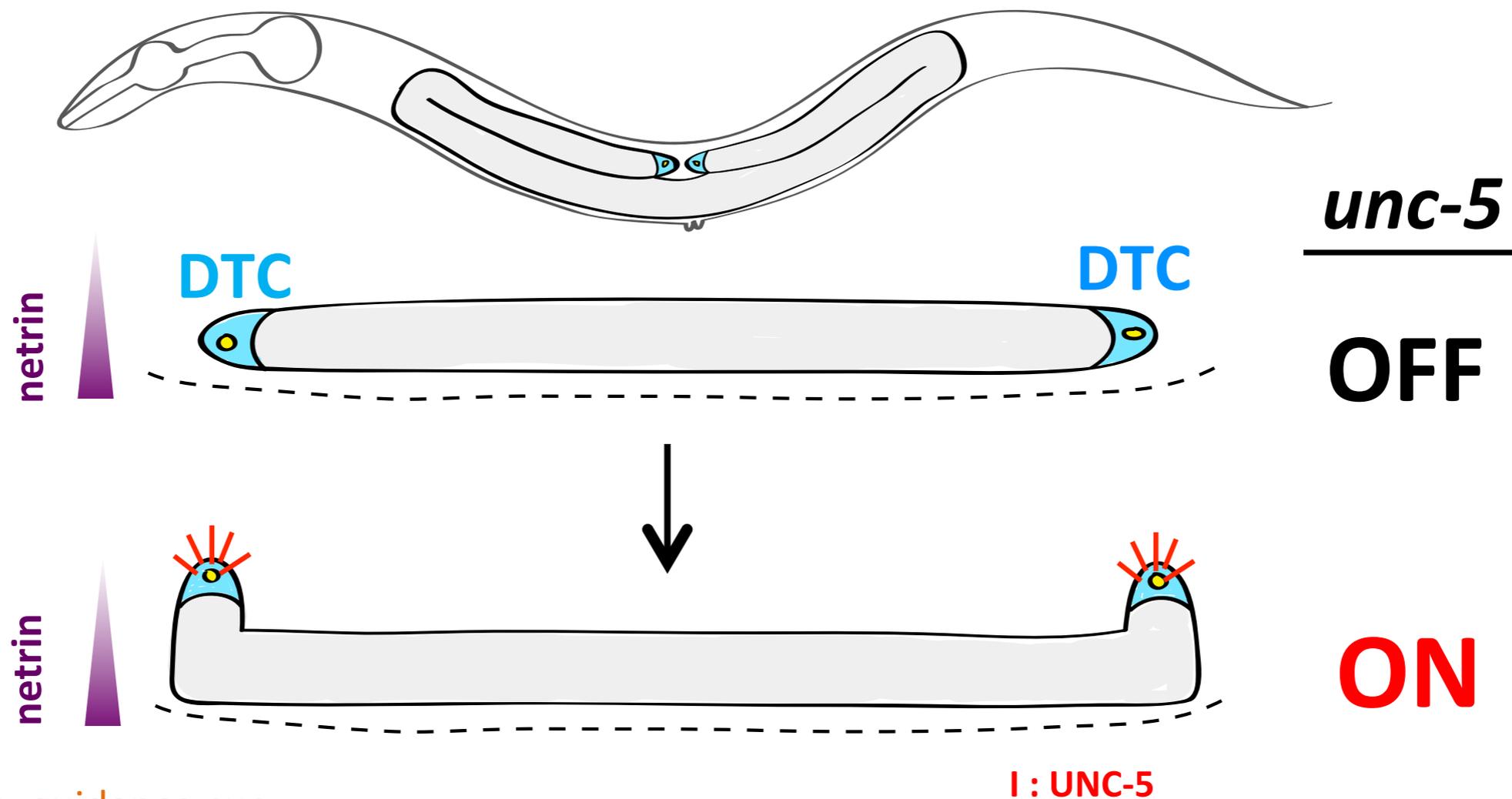
Collaboration project with NTU 吳益群教授

Noises in the development of worms (*C. elegans*)



Surendhar Reddy + Yi-Chen Chen 陳奕丞

The timing of gonad turn (*unc5* expression) is tightly



Genes Dev. 14, 2486-2500 (2000)

Development 127, 585-594 (2000)

Picture by courtesy of Yi-Ting Cheng 4

Is noise buffered by the gene regulation network?

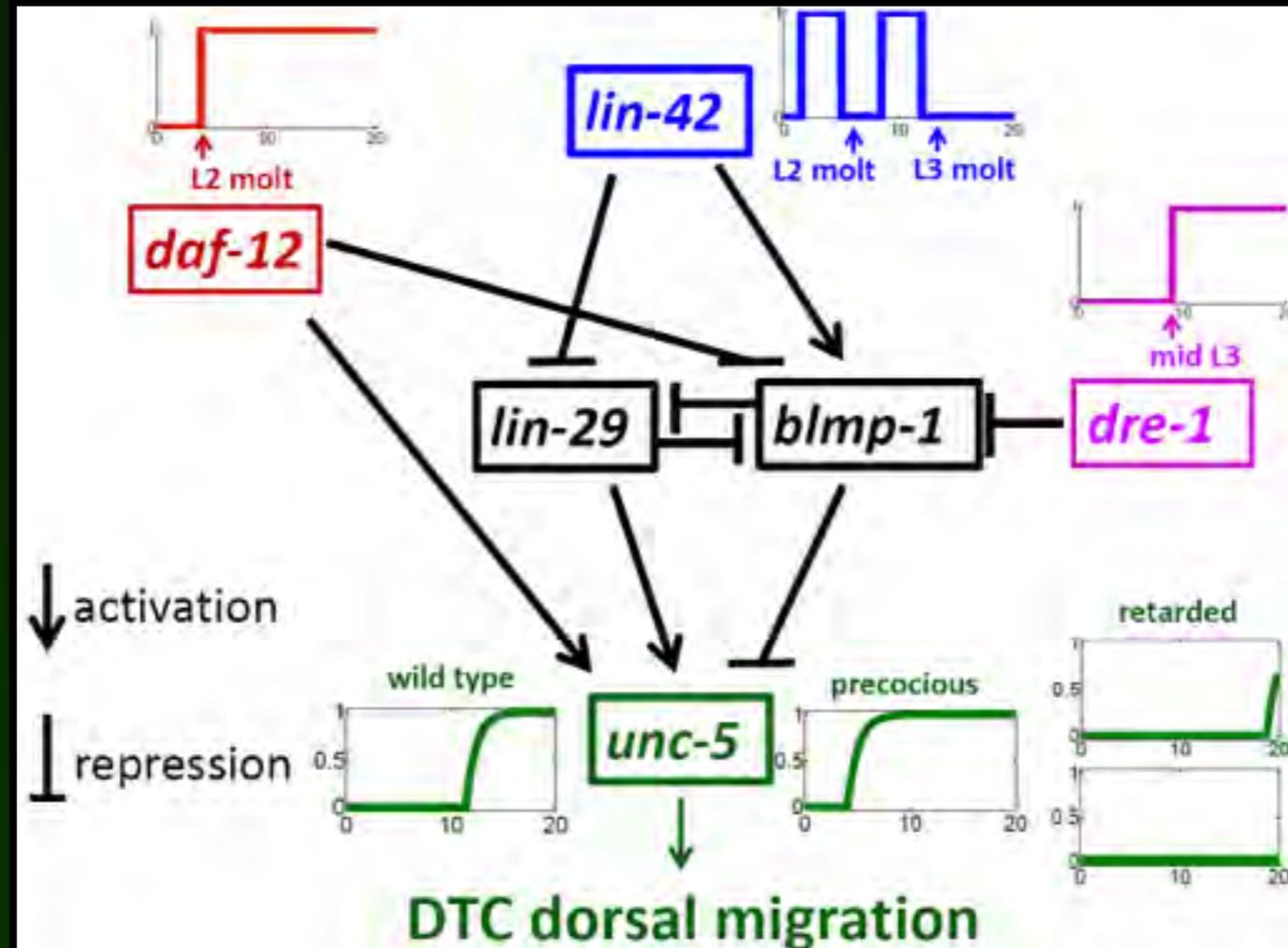
Phenotype: Dorsal turn
timing (and shape) %

GENOTYPE	WILD TYPE	PRECOCIOUS	RETARDED
<i>blmp-1(s71)</i>	7	93	0
<i>lin-29(n546)</i>	100	0	0
<i>lin-29(RNAi)</i>	100	0	0
<i>dre-1(dh99)</i>	100	0	0
<i>daf-12(rh61rh411)</i>	100	0	0
<i>lin-29(n546);dre-1(dh99)</i>	0	0	100
<i>lin-29(n546);daf-12(rh61rh411)</i>	0	0	97
<i>dre-1(dh99);daf-12(rh61rh411)</i>	0	0	98
<i>blmp-1(s71);daf-12(rh61rh411)</i>	43	47	10
<i>blmp-1(s71);lin-29(RNAi)</i>	30	54	16
<i>blmp-1(RNAi);lin-29(n546);dre-1(dh99)</i>	22	43	35
<i>blmp-1(s71);dre-1(dh99);daf-12(rh61rh411)</i>	12	73	15

Is there a noise-filtering mechanism?

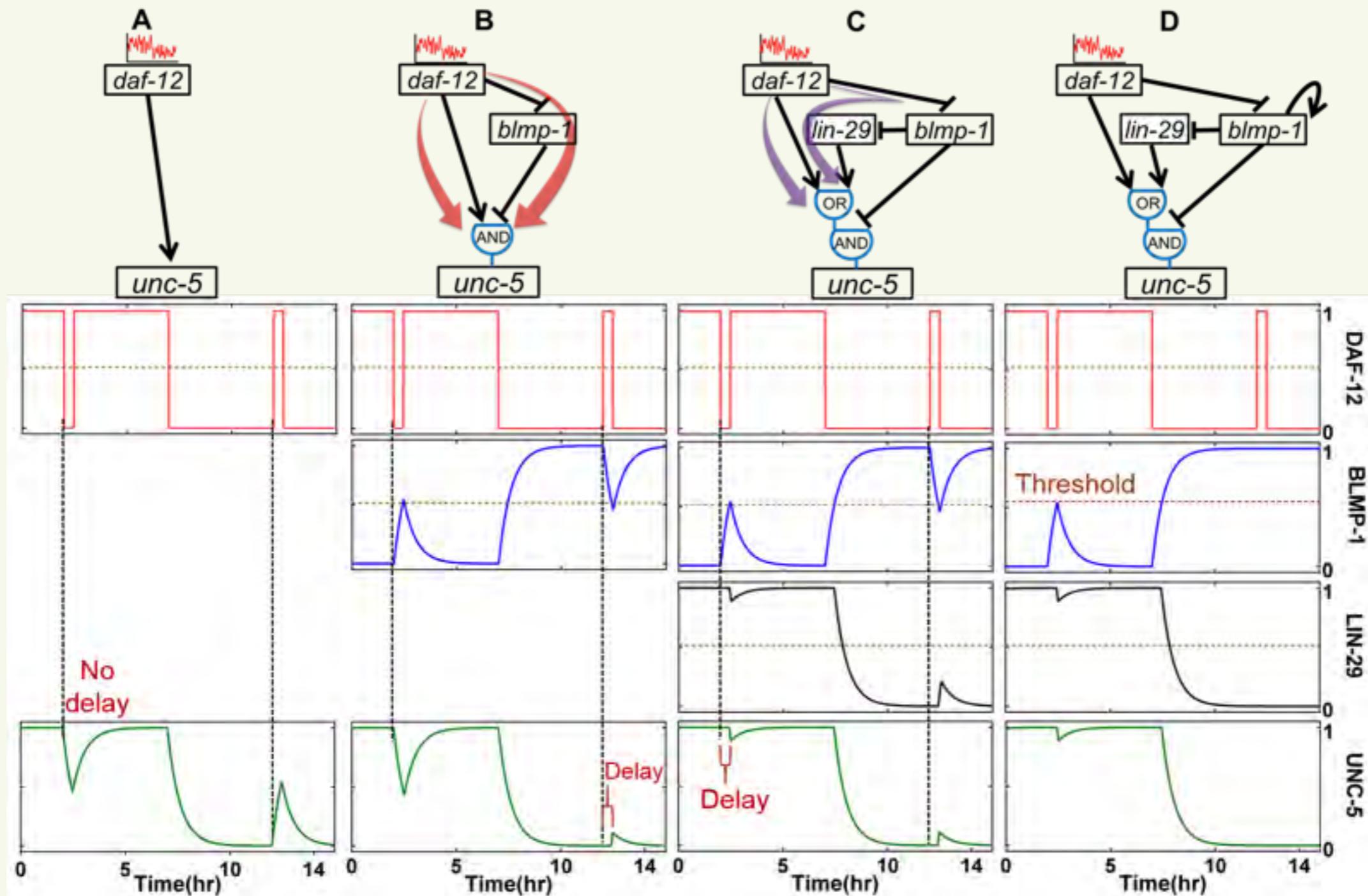
Lost of "noise-filtering" function?

We built a model that generates almost all experimental mutant phenotypes.



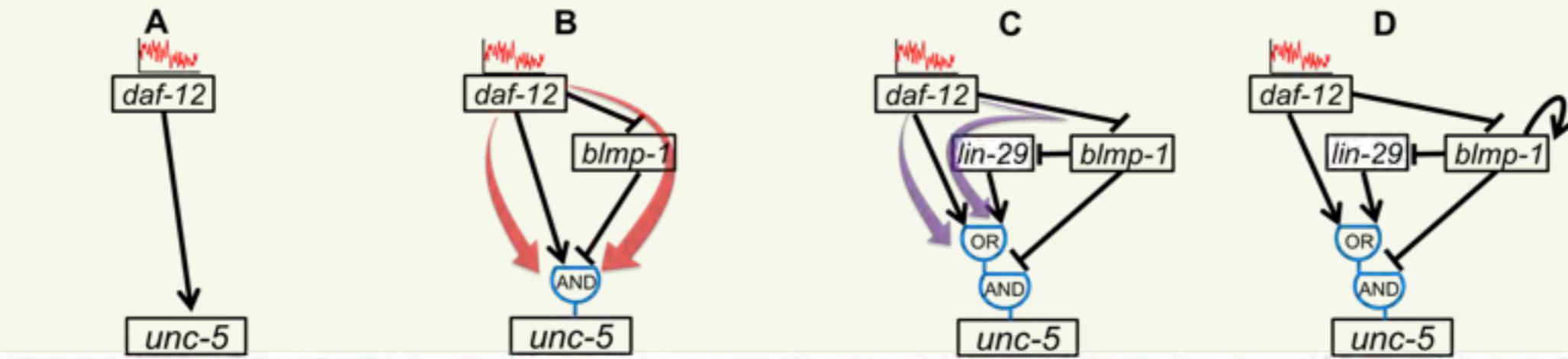
Node	Regulatory Logic
<i>lin-29</i>	$lin-29^* = \text{not } lin-42 \text{ and not } blmp-1$
<i>blmp-1</i>	$blmp-1^* = (lin-42 \text{ and not } lin-29 \text{ and not } daf-12) \text{ or } blmp-1$
<i>unc-5</i>	$unc5^* = (lin-29 \text{ or } daf-12) \text{ and not } blmp-1$

Interlinked FFL filters upstream noises



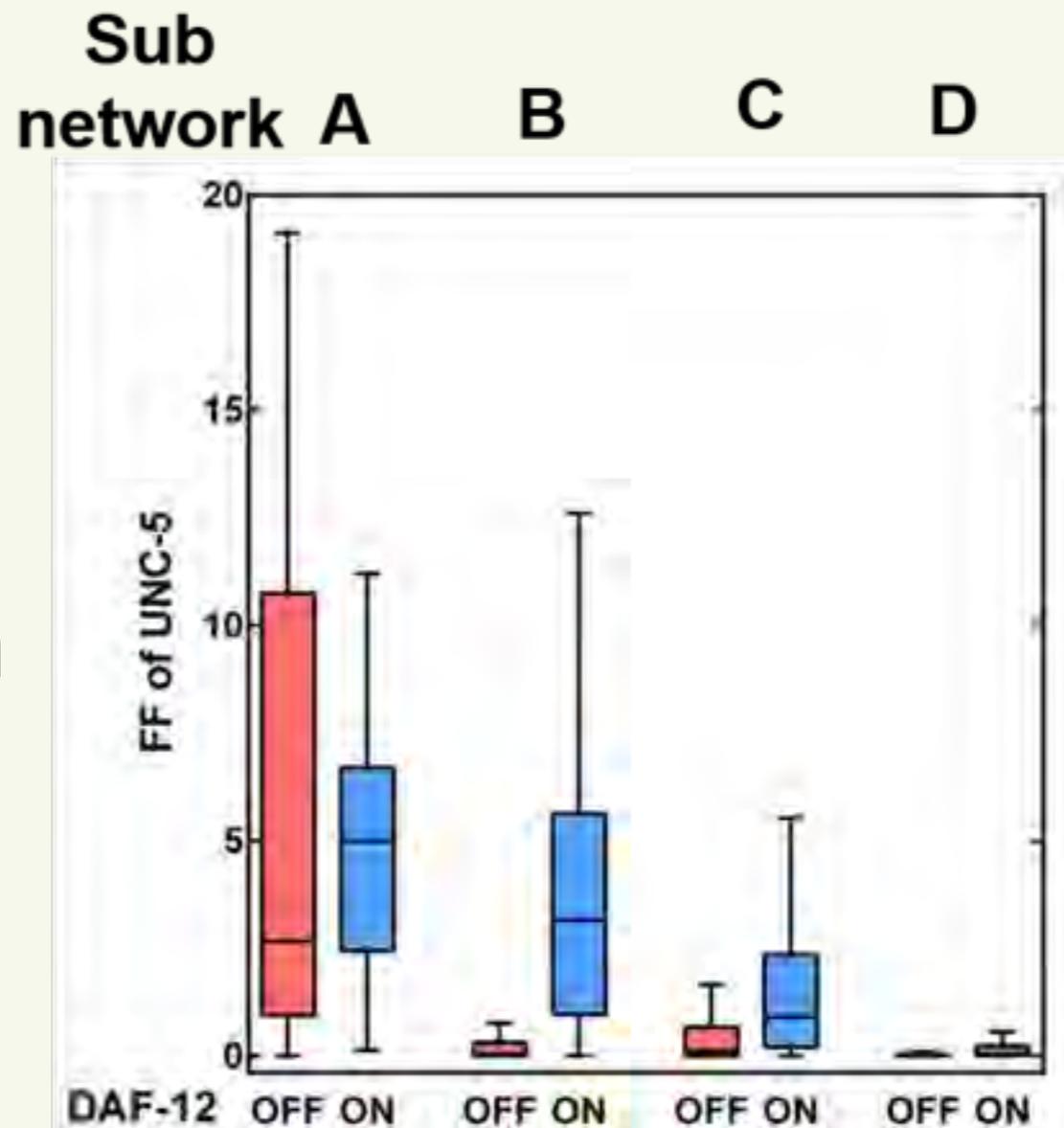
Adding noise in simulation

- Langevin's equation: Noise can be easily added and removed.
- All genes have "intrinsic" noises — the Gaussian noise in their production and degradation.
 - Production: burst noise considered.
- Regulated genes have propagated noise.
- Other noises (not considered) — Global noise.

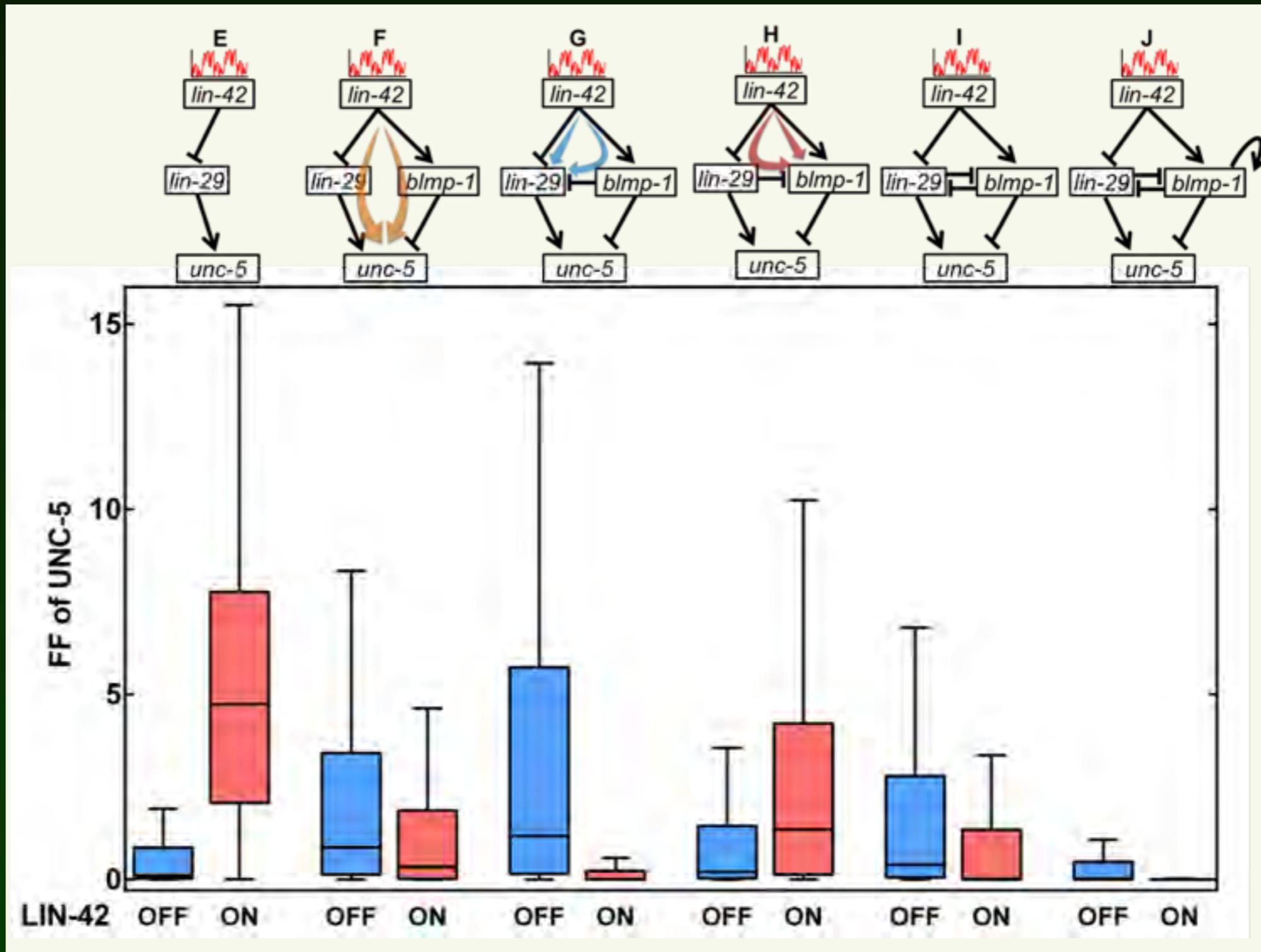


Fano Factor (FF)
= Variance/Mean

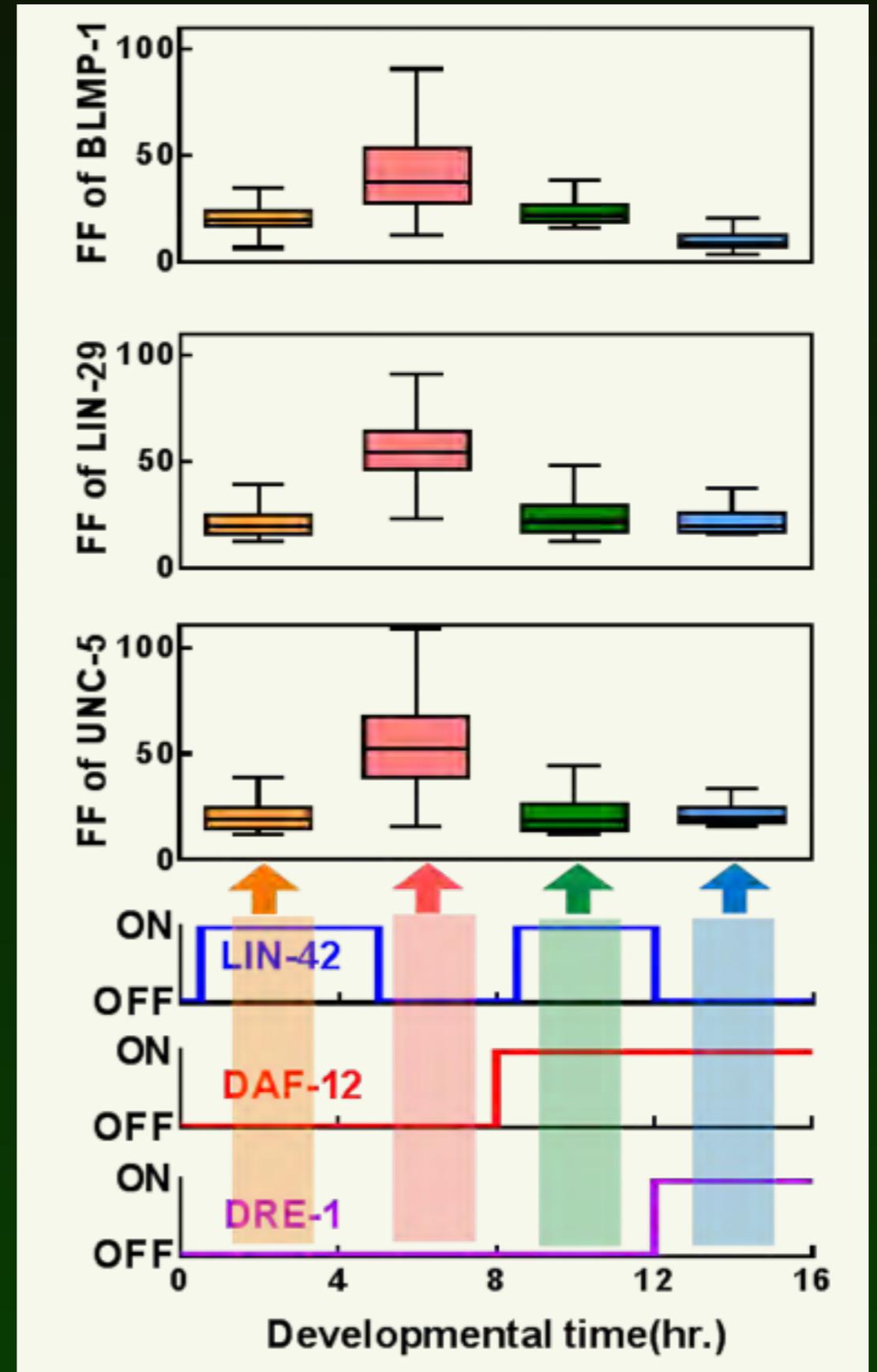
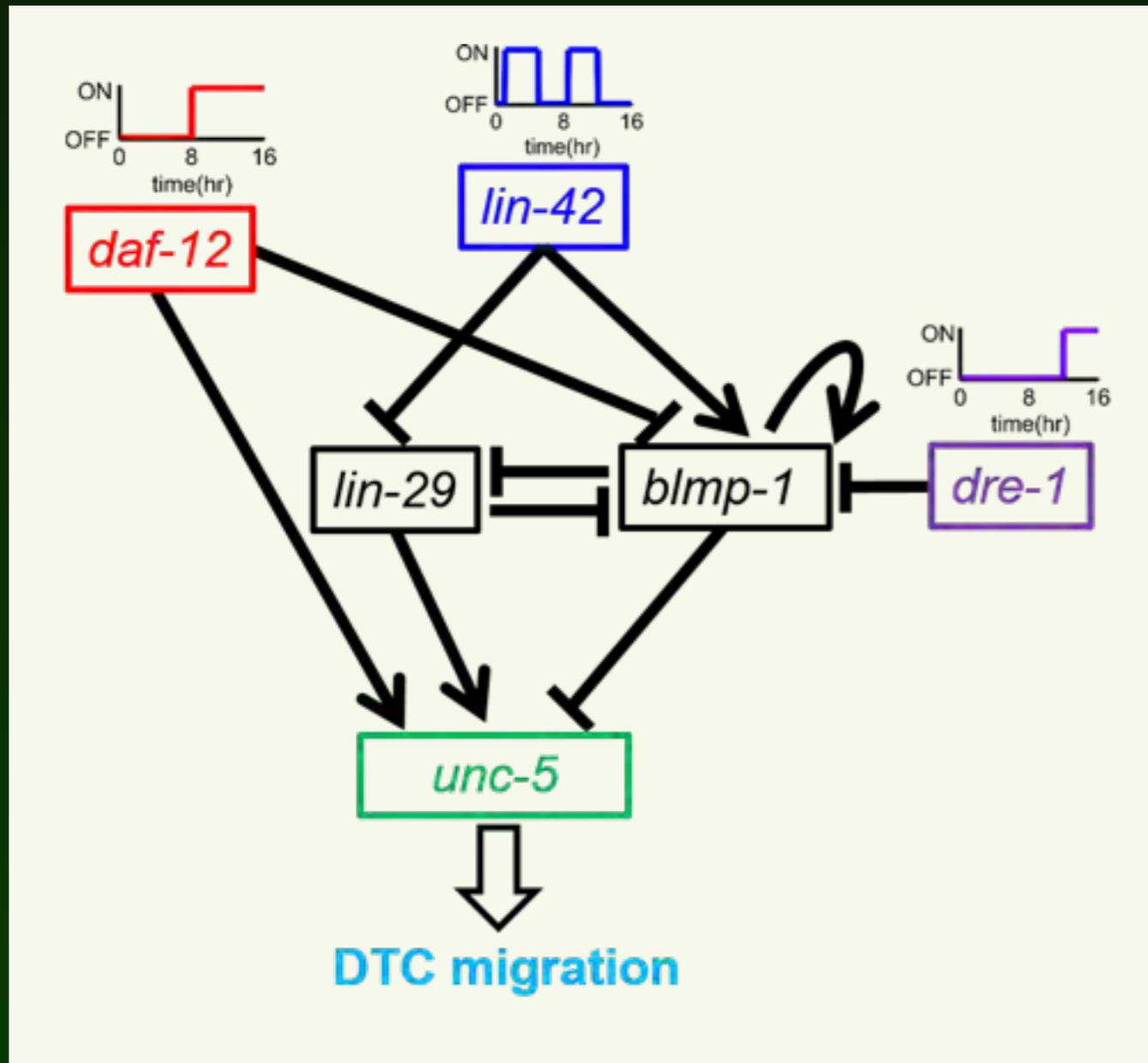
Results from 1000
randomly selected
parameter sets that can
produce wild type
phenotype.



Another set of IFFFL



All noise added & propagated

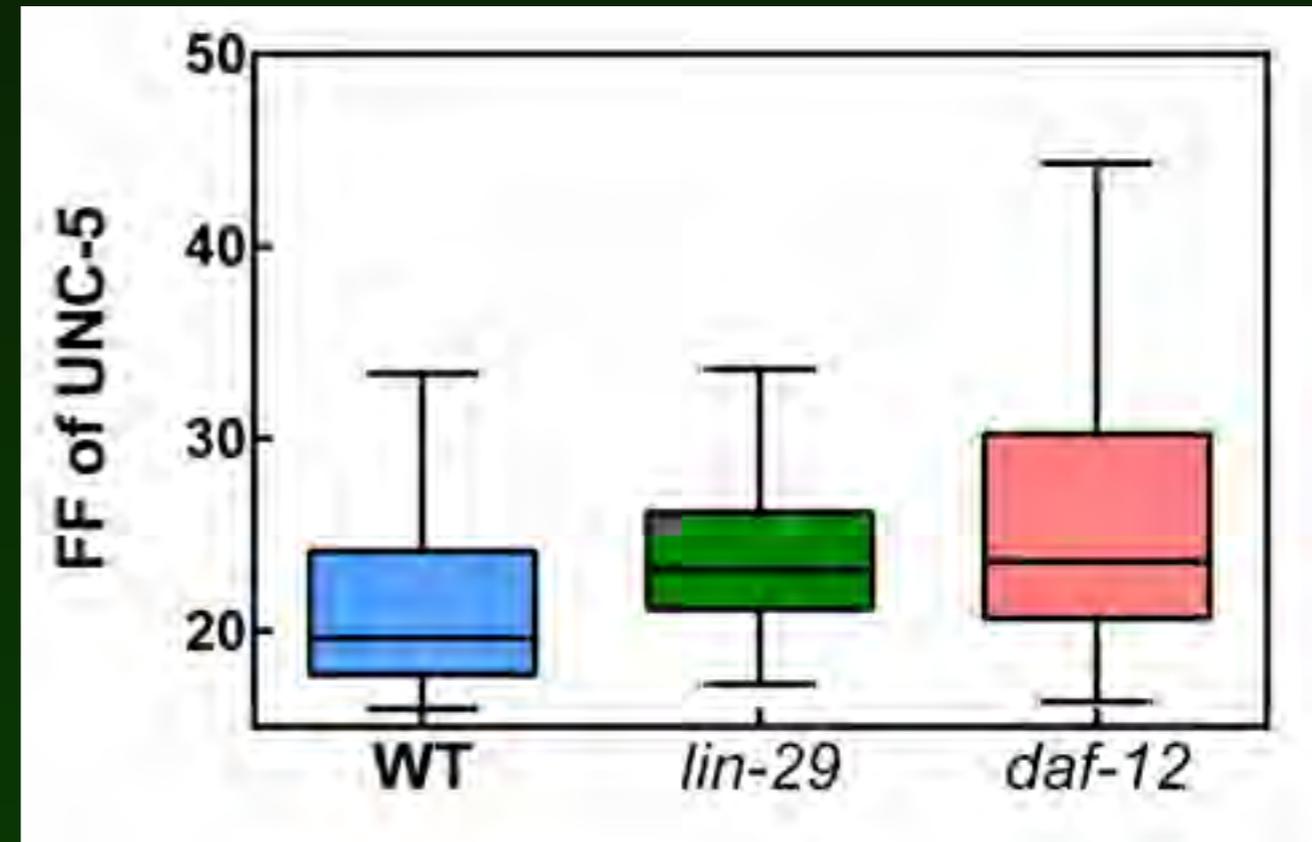
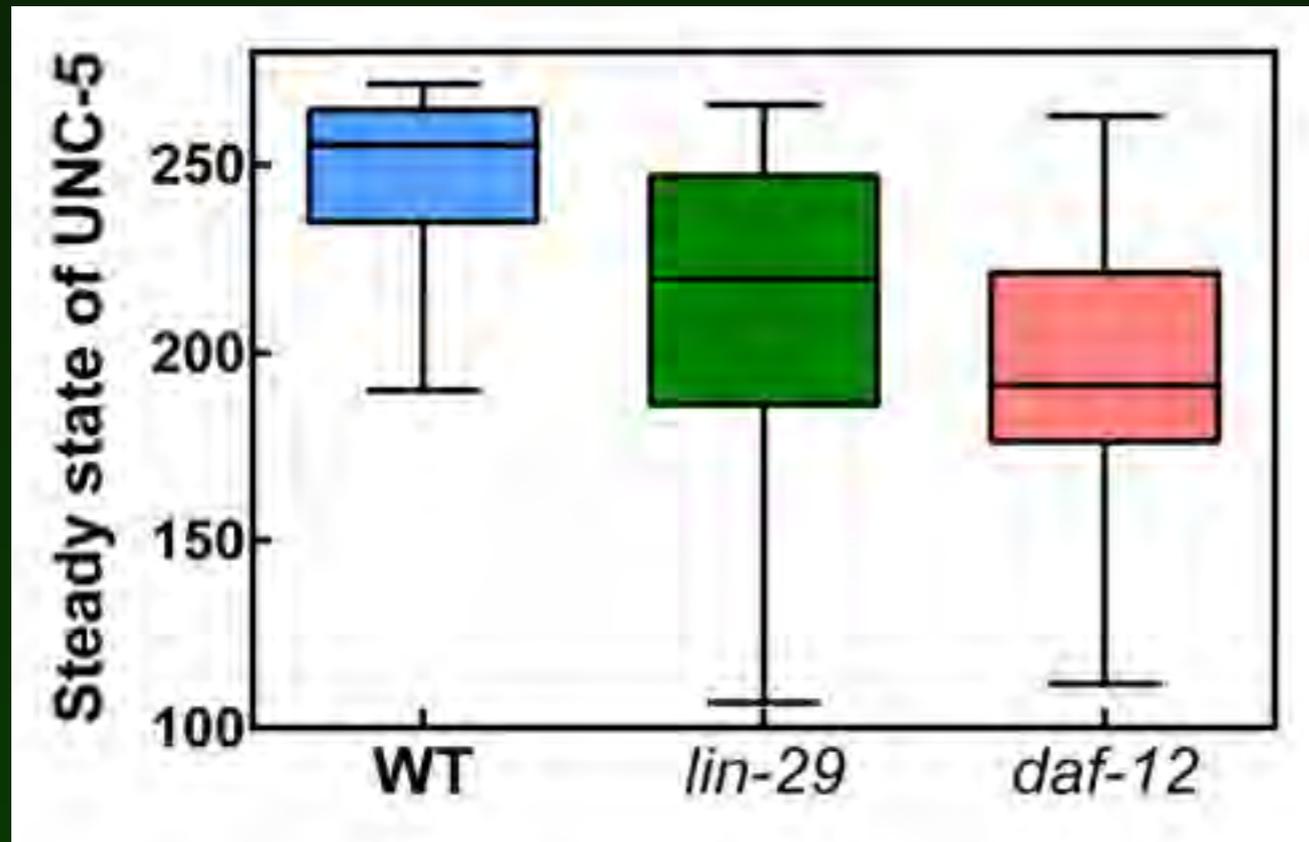
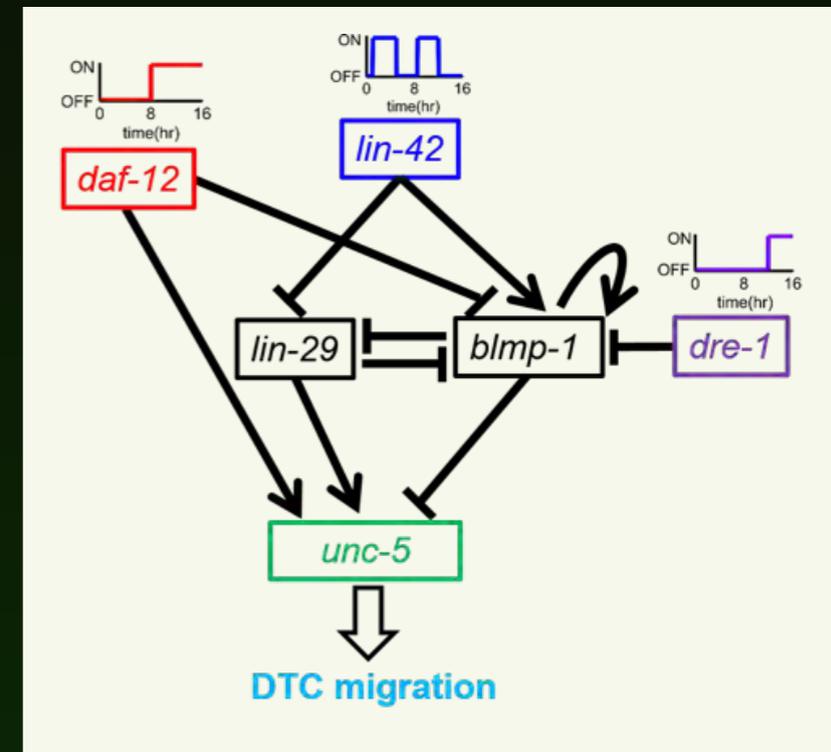


Is noise buffered by the gene regulation network?

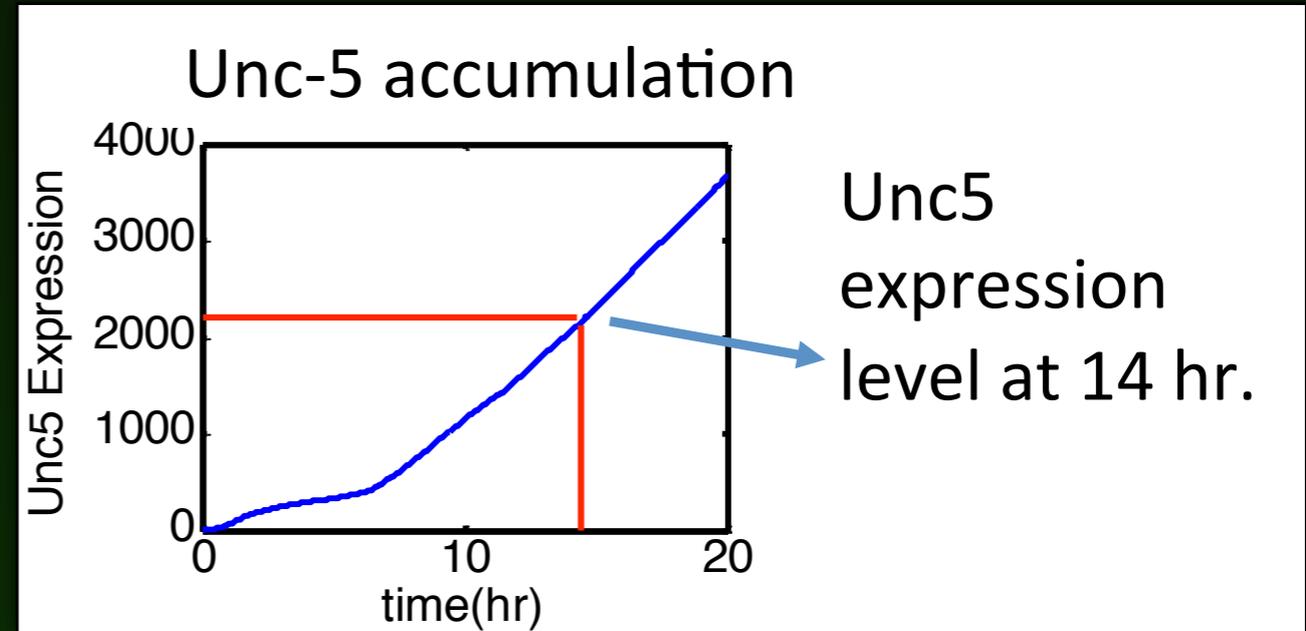
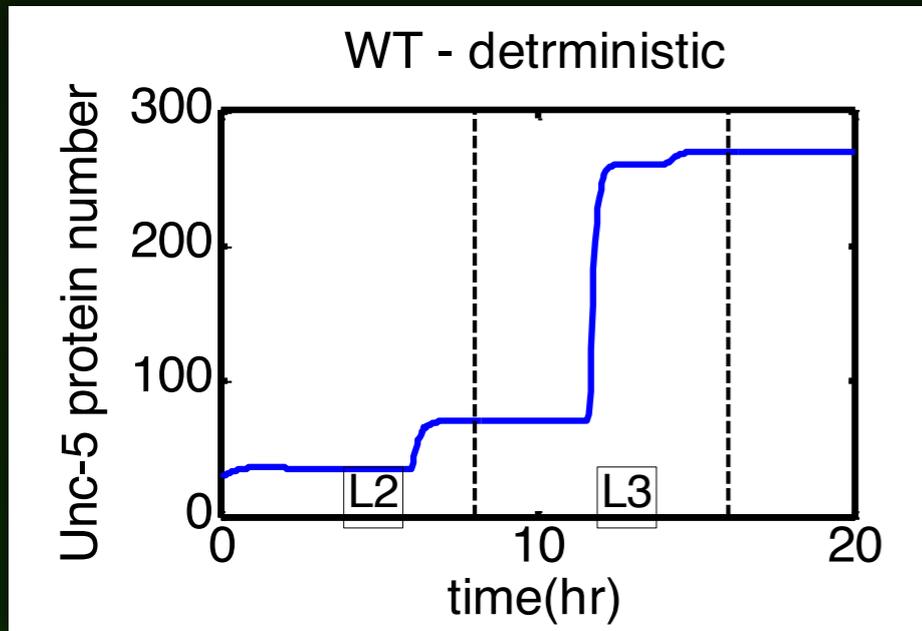
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<i>blmp-1(s71)</i>	7	93	0
<i>lin-29(n546)</i>	100	0	0
<i>lin-29(RNAi)</i>	100	0	0
<i>dre-1(dh99)</i>	100	0	0
<i>daf-12(rh61rh411)</i>	100	0	0
<i>lin-29(n546);dre-1(dh99)</i>	0	0	100
<i>lin-29(n546);daf-12(rh61rh411)</i>	0	0	97
<i>dre-1(dh99);daf-12(rh61rh411)</i>	0	0	98
<i>blmp-1(s71);daf-12(rh61rh411)</i>	43	47	10
<i>blmp-1(s71);lin-29(RNAi)</i>	30	54	16
<i>blmp-1(RNAi);lin-29(n546);dre-1(dh99)</i>	22	43	35
<i>blmp-1(s71);dre-1(dh99);daf-12(rh61rh411)</i>	12	73	15

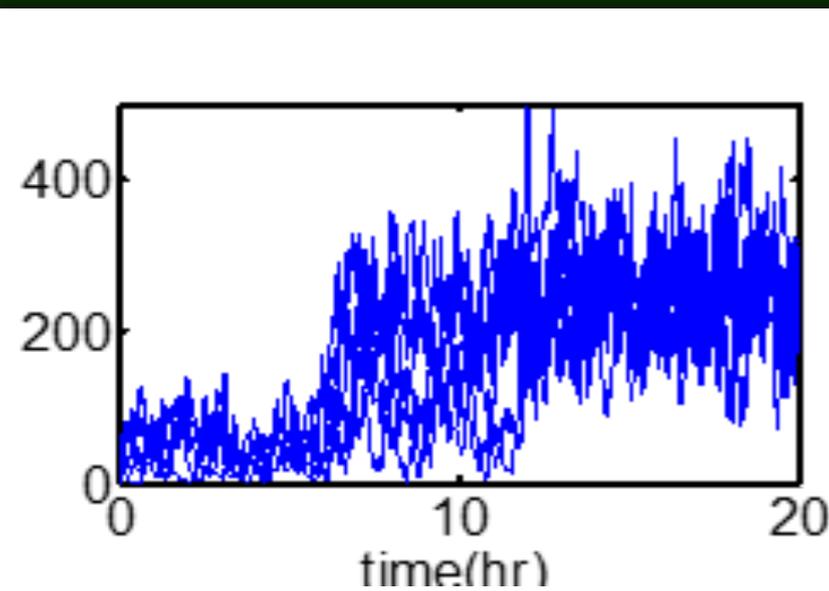
WT and Mutant behavior



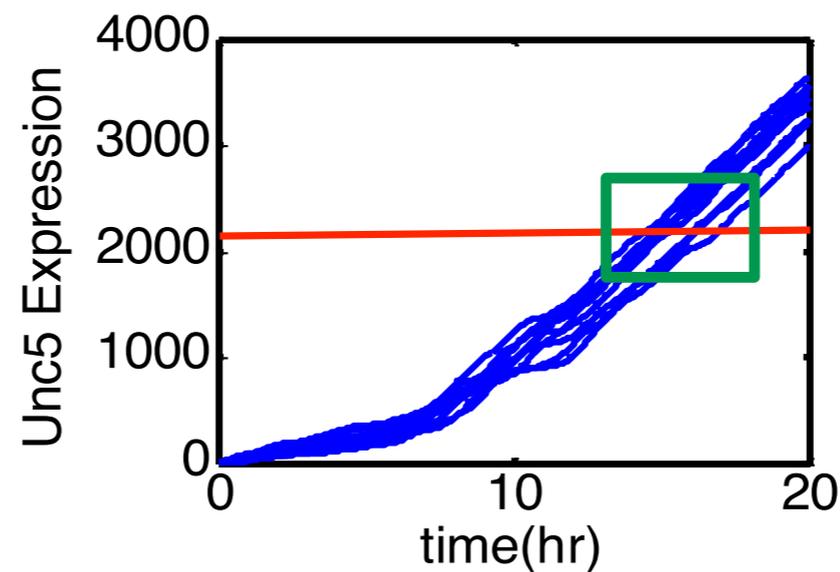
How shall we determine phenotype?



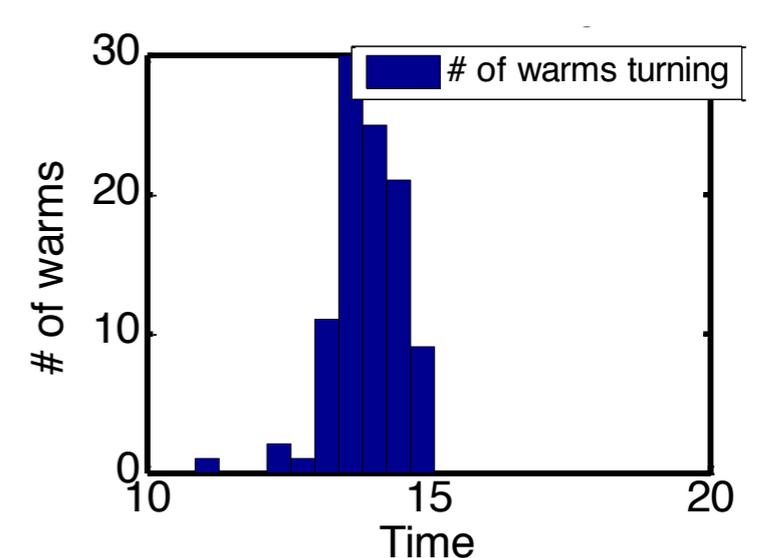
Stochastic unc-5 expression



Accumulated unc-5 signal



Time pass threshold



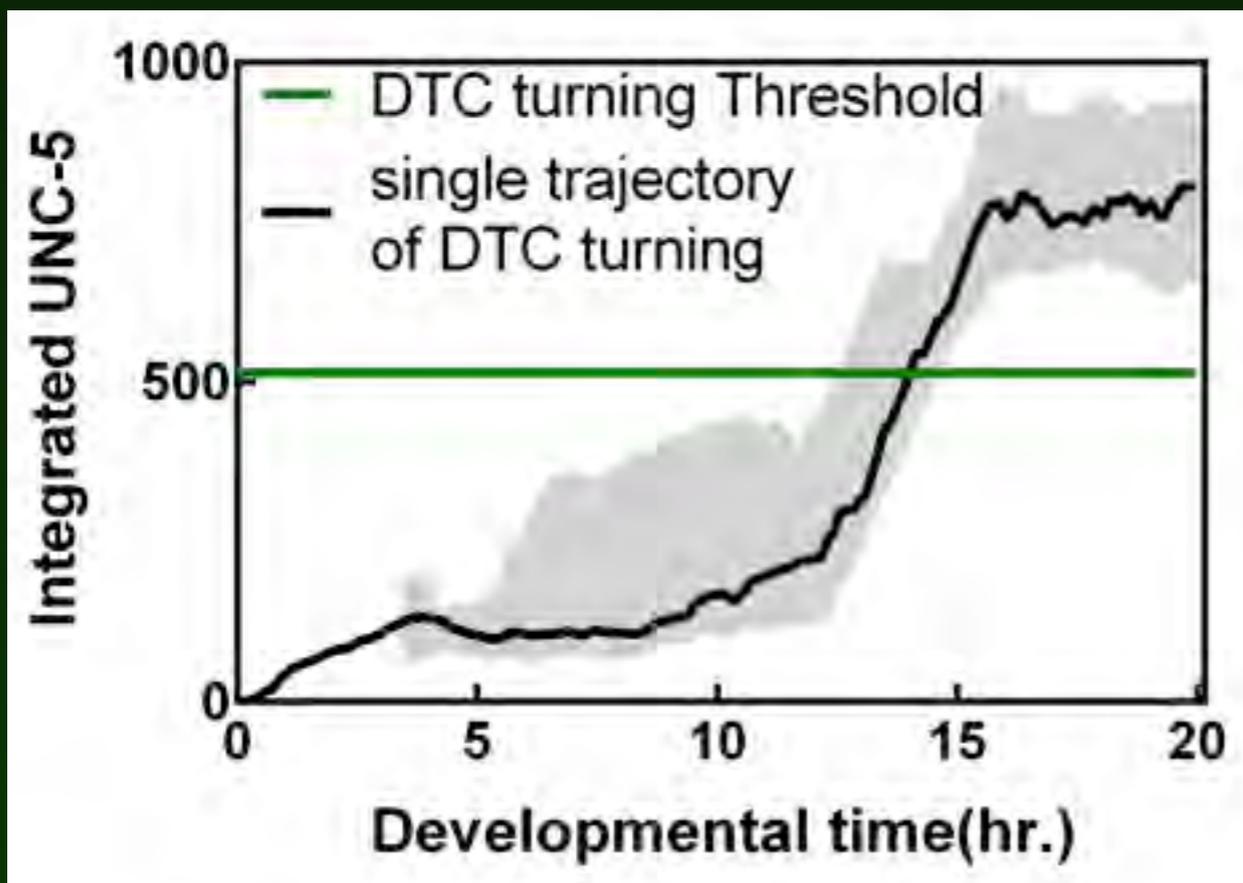
Is noise buffered by the gene regulation network?

Phenotype: Dorsal turn timing (and shape) %

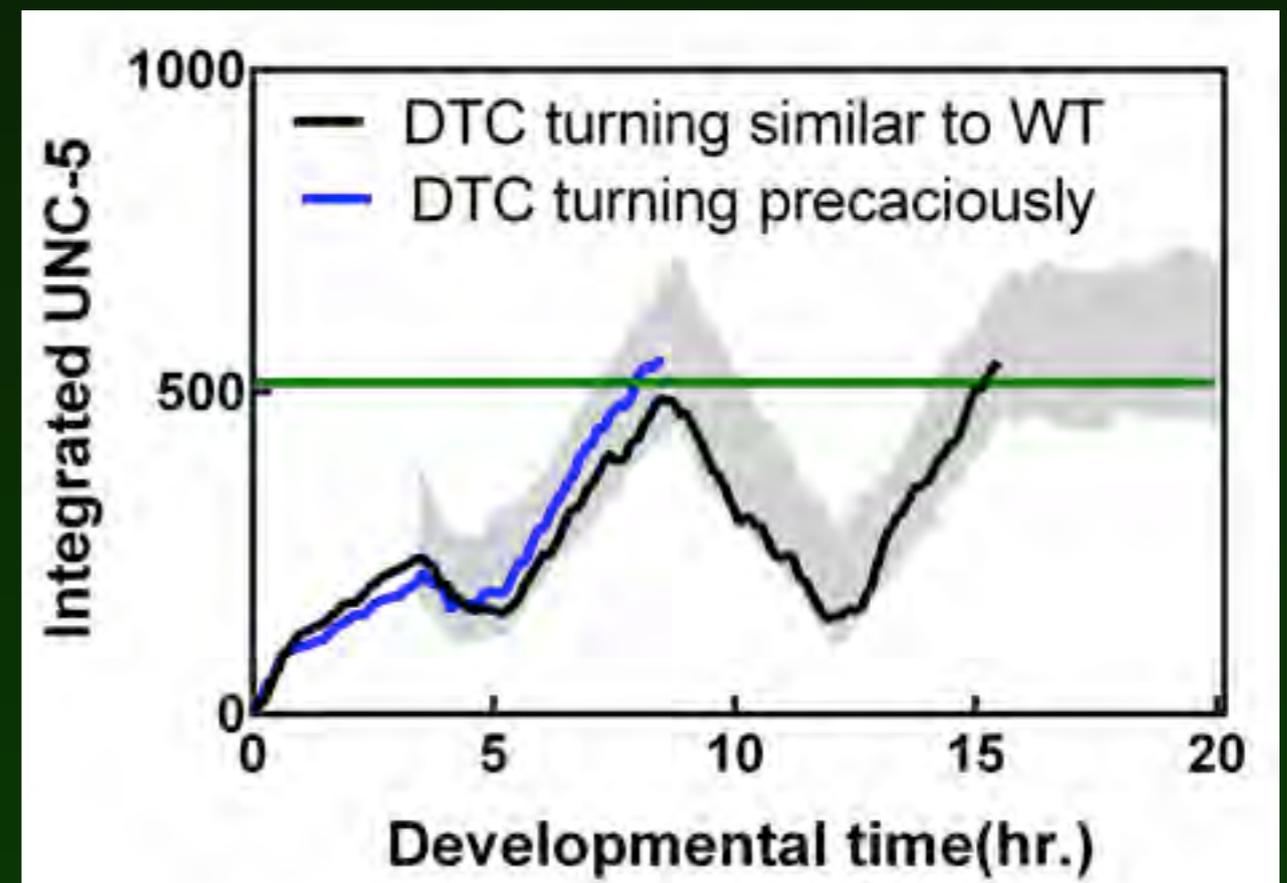
GENOTYPE	WILD TYPE	PRECOCIOUS	RETARDED
<i>blmp-1(s71)</i>	7	93	0
<i>lin-29(n546)</i>	100	0	0
<i>lin-29(RNAi)</i>	100	0	0
<i>dre-1(dh99)</i>	100	0	0
<i>daf-12(rh61rh411)</i>	100	0	0
<i>lin-29(n546);dre-1(dh99)</i>	0	0	100
<i>lin-29(n546);daf-12(rh61rh411)</i>	0	0	97
<i>dre-1(dh99);daf-12(rh61rh411)</i>	0	0	98
<i>blmp-1(s71);daf-12(rh61rh411)</i>	43	47	10
<i>blmp-1(s71);lin-29(RNAi)</i>	30	54	16
<i>blmp-1(RNAi);lin-29(n546);dre-1(dh99)</i>	22	43	35
<i>blmp-1(s71);dre-1(dh99);daf-12(rh61rh411)</i>	12	73	15

Simulating phenotypes

Wild type

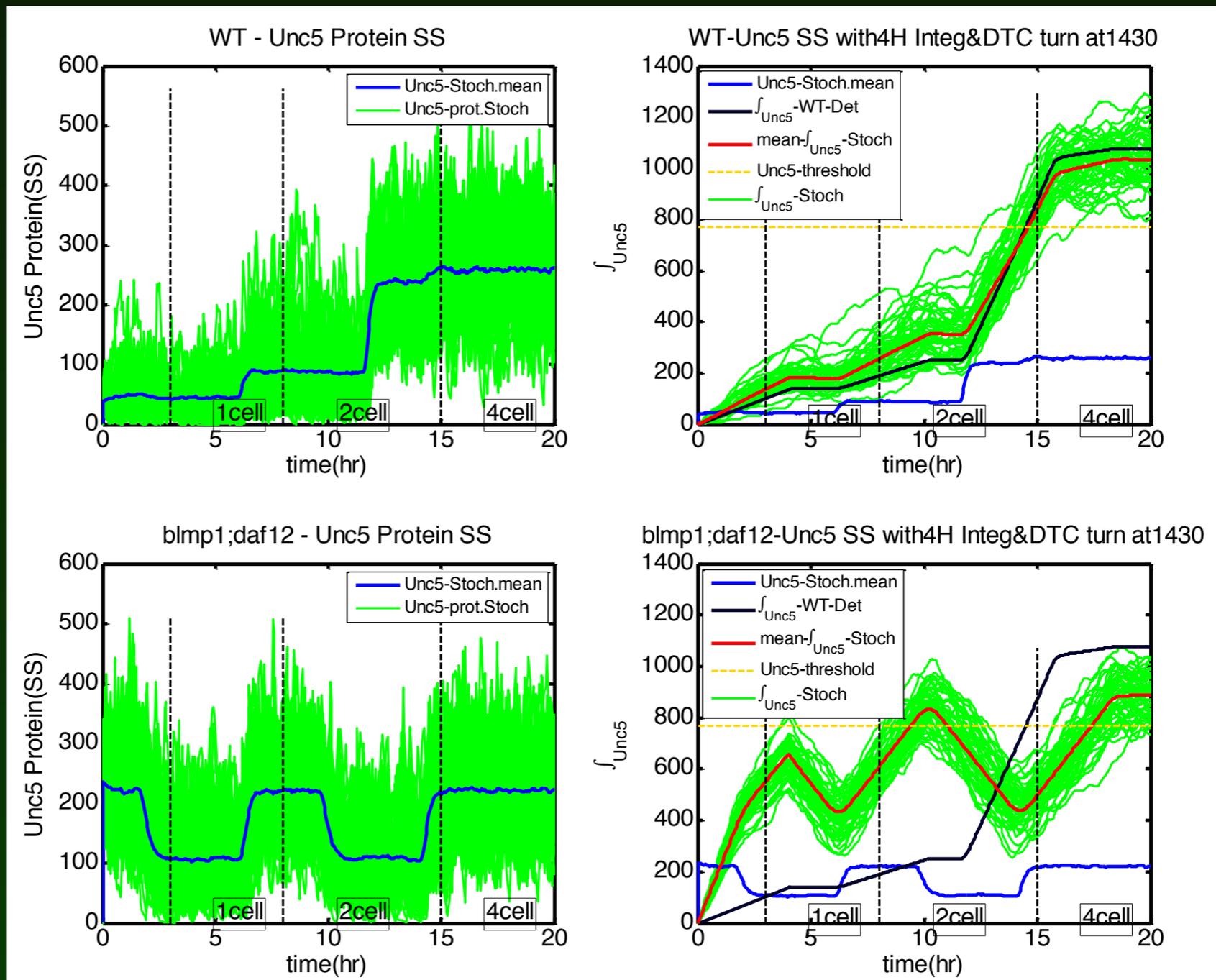


blmp-1 daf-12



Computer modeling helps us see why/how

WT



Mutant:
blimp1,daf12

Phenotype diversity

- Most previous work: gene expression noises \Rightarrow diverse phenotypes.
- Our result does not support such a link.
 - *unc5* expression uncertainty does not directly correlate with DTC turning phenotypes.
- It is [Dynamics + noises]
 - WT: stay low. Even with noisy expression phenotype remains uniform.
 - mutant: *unc5* expression goes up and down.

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